Fecal Analysis: Fecal samples were collected on during on days 184 and 185 (week 27) from fasted animals in control, LD, MD, HD and pair-fed control groups scheduled for terminal sacrifice and on day 186 from non-fasted animals in the same groups scheduled for recovery sacrifice. The samples were analyzed for fat, protein (% nitrogen), and pH. Dry weight of each sample was also recorded.

Hormone Analysis: Blood samples were collected from each animal in control, LD, MD, HD and pair-fed control (except those designated for PK analysis) at scheduled sacrifices. Serum harvested from the blood was analyzed for TSH, T4, T3, LH and PTH.

Vitamin D Analysis: Recovery animals were fasted overnight during week 28 and bled before recovery sacrifice. Serum harvested was analyzed for 1, 25-dihydroxyvitamin D.

Lymphocyte Subsets: To determine the immunotoxic potential of this compound, animals designated for recovery were fasted overnight during week 27 and whole blood was collected for lymphocyte subset analysis. A portion of spleen and thymus were also collected from animals in control, LD, MD, HD and pair-fed control at terminal sacrifice for lymphocyte subset analysis.

Results:

Mortality: No test material-related deaths. Sponsor stated that 2/30 (0 mg/kg), 2/40 (MD) and 1/48 (HD) died as a result of gavage error. Histopathology data was not provided.

Clinical signs: Incidence

Dose (mg/kg/d)		0	3(00	60	0	12	00		r fed itrol
n	30		48		48		48		30	
Sex	M	F	M	F	M	F	М	F	M	F
Swollen neck			1		2		2			
Swollen paws					2		3			
Scaly tail			1		3	10	22	22		
Scaly tail tip		1	5		4	5	4	3		2
Wart-like lesions on tail	1	1	1	7	5	4	11	8		1

Empty cells indicate zero incidence

Body weights: (g)

Table showing decrement and % decrement in mean body weight gain

Dose (mg/kg/d)	0		30	300		600		1200		fed trol
Sex	M	F	M	F	M	F	M	F	M	F
Day -4	125	122	124	119	124	119	124	118	124	115
Day 183	657	392	604*	416	580*	397	506*	356	592*	362
Decrement	0	0	53		. 77		151	36	65	30
% decrement	0	0	8	•	12	-	23	9	10	8
Day 218 (Recovery)	640	371	589	419	586	405	531*	341	599	348
Decrement	0	0	51		53	•	109	30	41	23
% decrement	0	0	8	•	10	•	17	8	6	6

*p < 0.05

Food consumption: (g)

Table showing decrement and % decrement in mean food consumption

Dose (mg/kg/d)	0		300		600		1200		Pair fed control	
Sex	M	F	M	F	M	F	М	F	M	F
Week -1	111	113	112	110	112	111	115	115	111	107
Week 26	184	138	176	154	170	151	160*	138	156*	129
Decrement	0	0	8	-	14	-	24	0	28	9
% decrement	0	0	4	•	- 8	•	13	0	15	7
Week 30 (Recovery)	178	141	188	161	180	158	174	137	169	128
Decrement	0	0	-		-	-	4	4	9	13
% decrement	0	0	-	-	•	· .	2	3	5	9

Ophthalmoscopy:

WEEK 26: SUMMARY OF OPHTHALMIC OBSERVATIONS

Dose (mg/kg/d)	1	D	30	00	6	00	12	00		r fed ntrol
Sex	M	F	M	F	M	F	M	F	M	F
Choroidal atrophy, OD			1/48		1/48	1/48	2/48			1/30
Dacryoadenitis, OS		3/30	1/48				2/48	1/48	1/30	1/30
Dacryoadenitis, OD			1/48							1/30
Corneal dystrophy, OD	1/30					l				
Corneal dystrophy,OS						1/48	1/48			
Corneal dystrophy,OU		1/30				1/48	2/48	1/48		
Anterior synechia, OS							1/48			
Choroidal hemorrhage, OS								1/48		
WE	EK 30: REC	OVERY (PHTHA	LMIC	DBSER\	ATION	S			
Inflammation of iris, OD			2/48							
Inflammation of cornea, OS		L					1/48			
Retinal hemorrhage, OD					l		1/48			
Chronic dacryoadenitis, OS		2/30			l —			1/48		
Chronic dacryoadenitis, OD		1/30								
Pthisis bulbi, OD		1/30						1/48		
Inflammation of iris, OD										1/30

Empty cells indicate zero incidence

Electrocardiography: No data.

Hematology:

WEEK 27 DATA

Dose (mg/kg/d)	0		300		600		1200		Pair fed control	
Sex	M	F	M	F	M	F	М	F	M	F
RBC (E6/UL)		7.8		7.5		7.3*		7.2*		7.7
HGB (g/dl)	15.4	15.5	14.7	14.7	14.6*	14.4*	14.7	14.0*	15.5	15.2
HCT (%)	1	42.8		41.1		40.3		39.2*		42.2
PLT (E3/UL)	1012	953	983	883	934	792*	787°	796°	981	883
WBC (E3/UL)		4.7		6.5		7.4*		8.0*		4.2
LYMPH (E3/UL)		4.0		5.6		5.7		6.5*		3.6
N-SEG (E3/UL)	1.5		2.2		6.0*		3.5*		1.4	

* p < 0.05

At the end of the recovery period (week 32) all hematology parameters had returned to normal limits.

Clinical chemistry:

WEEK 27 DATA

Dose (mg/kg/d)	0		300		600		1200		Pair fed control	
Sex	M	F	М	F	М	F	M	F	М	F
T. PRO (g/dl)	7.4		7.1		7.1		6.9*		7.1	1
ALB (g/dl)	4.8		4.3*		4.1*		4.1*		4.6	l
Bile acids (umol/l)	10		14		14		31*		14	L
AST (IU/L)	139		170		199		301*		148	
ALT (IU/L)	38		45		69*		114*		36	
ALKP (IU/L)	77		49*		59*		50°		77	
CHOL (mg/dl)		95		114		141 ± 59		134*		102
Electrophoresis ALB (g/dl)	5.0	± 17	4.6*	 	4.4*	1 59	4.5*	± 39	4.9	
α-1 globulin (g/dl)	0.2		0.5		0.7*		0.3		0.2	
γ-blobulin (g/dl)	0.4	0.6	0.2*	0.4 ± 0.10	0.2*	0.4 ± 0.21	0.3	0.3*	0.3	0.5
		W	EEK 31	(RECOVE	RY) DAT	Ά				
ALB (g/dl)	4.8		4.6		4.5*		4.3*		4.7	

	1 121	- 1 445 1	1 440	4040	120
I GLU (a/dl)	1 121 1	l 118 l	116	1 104° ł	
1 000 (9/0)					

SD was recorded in a few cases for clarification of statistical significance; * p < 0.05

Urinalysis:

WEEK 27 DATA

Dose (mg/kg/d)		0		300		600		00	Pair fed control	
Sex .	M	F	М	F	M	F	M	F	М	F
Ca EXC (mg)	1.1	2.2	1.8	4.6*	2.2°	5.2°	2.9*	4.4°	1.1	2.1
CI EXC (mmol)	0.5		0.7		C.8*		0.8*		0.5	
Urine Ca (mg/dl)		7.4		14.4*		14.0*		9.6		7.1
At the end of the recove	ry period (week 31) all uri	nalveis n	arameter	s had ret	urned to	normal li	mits	

EXC = excretion; * p < 0.05

Organ weights:

WEEK 27 DATA

Dose (mg/kg/d)	O		3	00	60	00	12	00	Pair con	
Sex	M	F	M	F	M	F	М	F	М	F
Epididymides (g)	1.56		1.24*		1.32*		1.15°		. 1.55	
Liver (g)	17.39		17.39		17.44		14.04*		14.23*	
Liver (%)	2.81	2.64	3.16*	3.15*	3.40*	3.33*	3.02	3.35*	2.63	:2.80
Heart (g)	1.76		1.67		1.51*		1.52*		1.58	١.
Spleen (g)	0.82		0.66		0.71		0.54*		0.65	
Stomach (g)	2.31		2.23		2.12		1.92*		2.04	
Pituitary (g)	0.017	0.024	0.018	0.037*	0.016	0.034*	0.015*	0.033	0.018	0.025
Pituitary (%)		0.006		0.010*		0.009*		0.010°		0.008
Small intestine (%)	1.42	2.04	1.64*	2.28	1.69*	2.35	1.85*	2.60*	1.47	2.17
Large intestine (%)	0.80	1.05	0.86	1.07	0.90*	1.17	0.94*	1.28*	0.80	1.03
Kidney (%)	0.62		0.68		0.75*		0.70		0.61	
Cecum (%)	0.230	0.306	0.258	0.341	0.261*	0.352	0.313*	0.408*	0.263	0.327
Brain (%)	0.37		0.41		0.44*		0.48*	<u> </u>	0.42*	
			WEE	K 32 (REC	OVERY D	ATA)				
Epididymides (g)	1.50		1.39		1.41		1.30*	l	1.51	
Pituitary (g)		0.025		0.031		0.032		0.033*		0.024
Pituitary (%)	0.0026	0.007	0.003 1*	0.008	0.003*	0.008	0.003*	0.011*	0.0028	0.007 4
Small intestine (%)	1.25		1.39		1.51*		1.49*		1.27	
Large intestine (%)	0.73		0.750		0.80		0.87*		0.72	
Kidney (%)	0.60		0.68		0.65		0.71*		0.60	
Brain (%)	0.38		0.40		0.40		0.44*		0.40	

Absolute wt (g); Relative wt (%) = relative to body wt; * p < 0.05

Gross pathology:

WEEK 27 DATA

Dose (mg/kg/d)	()	3(00	60	0	12	00	Pair fed	control
Sex	М	F	M	F	M	F	M	F	M	F
Kidneys Large pelvis(es)					1/10		1/10			
Kidneys Cysts						_		1/10	,	
		 	WEE	32 (RE	COVERY D	ATA)				
Epididymides Small							1/10			
Testes Small			1/10				1/10			

Empty cells indicate zero incidence

Histopathology: Please note that sponsor did not provide severity scores for some lesions.

INTERIM SACRIFICE (WEEK 14)

Dose (mg/kg/d)	0		300		600		1200		Pair fe contro	
Sex	M	F	M	F	M	F	M	F	M	F
Pancreas - apoptosis Individual cell necrosis							1/5(1)	1/5(1)		
Liver Coagulative necrosis							1/5			
Stomach - Chief cells Cytoplasmic vacuolation							1/5(1)	1/5(1)		
Mucosal necrosis					1/5(2)					
Mesenteric L. node Lymphoid depletion							1/5			
Lungs Alveolar histiocytosis								1/5		
Kidney Mineralization, tubular		1/5(1)			•			3/5(1)		
Renal tubular dilatation								1/5(1)		

1 = slight; 2 = mild; 3 = moderate; 4 = marked; 5 = severe

TERMINAL SACRIFICE (WEEK 27)

Dose (mg/kg/d)		0	3	00	6	00	12	200		fed trol\
Sex	M	F	M	F	M	F	M	F	M	F
Esophagus Fibrosis, serosal							1/15	1/14		
Large intestine Mucosal necrosis							1/15			
Pancreas Acinar cell vacuolation							1/15(1)			
Salivary gland Cytokaryomegaly, seromucus acinar cells							1/15			
Stomach, chief cells Cytoplasmic vacuolation							2/15 1/15(1) 1/15(2)			
Epididymides Hypospermia							3/15			
Testis Degeneration/atrophy			1/15(1)		1/15(4)		3/15 1/15(2) 2/15(3)		1/15 (4)	
Mesenteric L. node Lymphoid depletion							1/15			
Skin, tail Hyperkeratosis, acantho -sis, pyogranulomatous dermatitis							8/15	,		
Eye Posterior synechia				l.			1/15			
Caratact, sub-capsular						l	1/15			
Kidney Chronic progressive nephropathy	3/13 1/13(1) 2/13(2)	1/15(1)	13/15 7/15(1) 5/15(2) 1/15(3)	8/15 5/15(1) 3/15(2)	12/15 5/15(1) 5/15(2) 2/15(3)	9/15 5/15(1) 3/15(2) 1/15(3)	12/15 4/15(1) 4/15(2) 1/15(3)	4/14 3/15(1) 1/15(2)		1/15 (1)

1 = slight; 2 = mild; 3 = moderate; 4 = marked; 5 = severe

RECOVERY SACRIFICE (WEEK 32)

Dose (mg/kg/d)	0		300	,	60	00	12	00	Pair con	
Sex	M	F	M	F	M	F	M	F	M	F
Testes	1									
Degeneration/atrophy	1		1/10(4)			<u> </u>	2/10(4)			
Kidney							6/10			
Chronic progressive	2/9(1)				1	.	3/10(1)	1/10(1)		

nephropathy		 I		3/10(2)		T	 İ
Interstitial nephritis		 ļ — —			1/10(1)		

1 = slight; 2 = mild; 3 = moderate; 4 = marked; 5 = severe

Sperm evaluation:

WEEK 27: SUMMARY OF SPERM EVALUATION

Dose (mg/kg/d)	0	300	600	1200	Pair fed control
Sperm motility		1			
Total motile sperm	170 ± 69	59 ± 55	79 ± 62	50 ± 59	157 ± 63
Percent motility	41 ± 11	23 ± 16	27 ± 15	18 ± 18	42 ± 10
Sperm concentration (10°)	11 ± 3	6 ± 2	8 ± 5	5 ± 3	12 ± 3
Sperm Morphology					
Normal	98 ± 2	60 ± 21	75 ± 23	55 ± 28	98 ± 2
Amorphous	0 ± 0	3 ± 3	2 ± 4	10 ± 26	0.1 ± 0.4
Decapitated head	2 ± 2	36 ± 20	23 ± 22	33 ± 22	1 ± 2
Abnormal sperm (%)	2 ± 2	40 ± 21	26 ± 23	45 ± 28	2 ± 2
WEEK 3	2 (RECOVERY): SU	JMMARY OF SE	ERM EVALUA	TION	
Sperm motility	<u> </u>	1			1
Total motile sperm	162 ± 60	133 ± 38	138 ± 65	91 ± 74	201 ± 49
Percent motility	45 ± 7	45 ± 6	42 ± 11	31 ± 22	49 ± 5
Sperm concentration (10°)	12 ± 4	8 ± 3	7 ± 2	5 ± 4	12 ± 2 .
Sperm Morphology				I	
Normal	98 ± 2	88 ± 9	86 ± 13	64 ± 36	98 ± 3
Amorphous	0 ± 0	1 ± 2	0.1 ± 0.3	2 ± 3	0.1 ± 0.3
Decapitated head	2 ± 2	10 ± 10	10 ± 11	26 ± 29	2 ± 3
Abnormal sperm (%)	2 ± 2	12 ± 9	14 ± 13	36 ± 36	3 ± 3

Hormone Analysis:

WEEK 27 DATA

Dose (mg/kg/	d)		0	30	00	6	00	. 12	00	Pair con	
Sex		М	F	M	F	M	F	M	F	М	F
Thyroxine (µg/dl)	М	4.2	2.2	3.1*	2.0	2.4°	2.0	3.2*	2.8*	3.3	2.2
, , ,	SD	0.56	0.40	1.007	0.51	0.83	0.56	1.07	0.55	0.79	0.54
Triiodothyronine (ng	/ml) M	73.4	84.4	62.4	85.7	61.6	80.2	51.5°	90.2	55.5*	80.6
, ,	SD] 11]	15	19	14	12	18	8	18	12	26
			WE	EK 32 (RI	COVERY	Y DATA)					

At the end of the recovery period, T3 and T4 in treated males were not significantly different from controls. No T3 and T4 data was provided for recovery females.

Bone Mineral analysis:

WEEK 27 DATA

Dose (mg/kg/d)	0)	30	0	60	0	120	00	Pair con	
Sex	M	F	M	F	M	F	M	F	M	F
Dry Bone wt (g)	0.82		0.74*		0.71**		0.66*		0.75	

Dry bone weight of treated females were not statistically significantly different from control. None of the bone minerals analyzed (Ca, P) were statistically significantly different from control.

* p < 0.05

Fecal Analysis:

WFFK 27 DATA

Dose (mg/kg/d))	3	00	60	0	12	00	Pair con	
Sex	M	F	. M	F	M	F	М	F	М	F
Nitrogen (%)	3.70	3.74	3.81	3.74	4.05°	3.78	4.17*	3.89	4.32°	4.32°
Fat (%)	4.6		4.9		4.9		5.0°		5.0°	

^{*} p < 0.05

Serum 1, 25-Dihydroxyvitamin D (pg/ml):

WEEK 28 DATA

Dose (mg/kg/d)	()	3	00	6	000	12	500		r fed ntrol
Sex	M	F	M	F	M	F	M	F	М	F
1, 25-Dihydroxyvitamin D		11.5		7.66		6.33*		7.44		11.6
1, 25-Dihydroxyvitamin D was achieve statistical significance		y signific	antly de	creased o	only in N		s. Diffe		males	

* p < 0.05

Lymphocyte subsets:

Lymphocyte Subsets (Absolute Numbers) in Rat Peripheral Whole Blood Absolute Cell Number (X10³)/mm³ of Blood (MEAN ± SD)

	FE	(ALE_	
LYMPHOCYTE SUBSETS	CONTROL (p=3)	PAIR-FED CONTROL (9-9)	1200 mg/kg/d (n=8)
Total Lymphocyscs	41±19	44±12	60±20
CDASKA"	1.4 ± 0.9	13±04	21±09
PerT	24 ± 1.0	27±08	3.4 ± 1.3
C03+	23±09	27±08	3.4 ± 1.1
CD3+CD4+	15206	13:06	26 2080
CDS-CDS-	108+04	09+04	08+03

	M	ALE.	
LYMPHOCYTE SUBSETS	CONTROL (3-3)	PAIR-FED CONTROL (s-8)	1200 mg/kg/d (n=9)
Total Lymphocytes	5.9 ± 1.9	6.4 ± 1.9	ស±2#
CD45RA*	2.1 ± 0.7	24±0.7	3.6 ± 1.1°
Pan T	3.1 ± 1.2	3.3 ± 1.1	3.7 ± 1.6
CD3	3.0 ± 1.2	33±1.1	36116
CD3+CD4°	1.8 ± 0.7	2.0 ± 0.4	26 ± 1.0
CD3+CD&	13 ± 0.6	1.3 ± 0.7	1.4 ± 0.6

Absolute cell numbers were calculated by multiplying the "ourrected" lymphocyte subset percentages by the absolute hymphocyte cell count.

, - br 0'02

Abbreviations and Symbols: n - The number of enimals included in analysis

Lymphocyte Subsets (Absolute Numbers) In Rat Spieen Absolute Cell Number (X107)/Spleen (MEAN ± SD)^a

			PMALE		
LYMPHOCYTE SUBSETS	CONTROL	PAIR-FED CONTROL (9-5)	300 mg/kg/d (n=5)	600 mg/kg/d (s=5)	1200 mg/kg/d (tr-5)
Total , Leukocytes	23.9 2.5.2	146233*	21.7 ± 7.1	262±143	19.6 ± 10.2
CD45:	93±23	5.5 ± 2.0°	9.7 ± 3.2	88:45	78±53
cm.	10.7 ± 1.7	65±16-	10.8 ± 3.4	10.7 ± 5.5	98 ± 6.2
CON CONT	54 ± 1.5	34±10*	6.7±25	6.4±3.2	61±41
concor	42±03	2.2 ± 0.6°	3.4±0.8	3.1 ± 1.7	2.2 ± 1.4
lg)AF	11.7 ± 2.4	6.6 2 1.4"	9.7 ± 4.0	11.9±7.2	8.9 ± 4.7

			MALE		
LYMPHOCYTE SUBSETS	CONTROL (==0)	PAIR-FED CONTROL (=-5)	300 mg/kg/d (s=3)	600 explicate (a=5)	1200 mg/kg/d (p=5)
Total Leukocytes	23.4±7.9	21.4 ± 9.6	26.7 ± 13.6	21.7 ± 1.8	22.0 : 8.0
CD42.	8.4 2 2.5	69±16	11.2 ± 5.8	7.7 ± 1.3	8.1 ± 3.3
CD3.	9.7±3.5	9.4±3.4	12.7 ± 6.2	9.6 ± 1.6	10.2 ± 4.4
ന്നാന	5.8 ± 2.1	5.2±0.8	7.7 ± 5.4	65 ± 2.0	68±3
CD3.CD8.	33213	29±12	3.5 ± 1.9	21±02	24±07
FM	11.8 ± 4.1	119113	11.0 ± 5.6	10.3 ± 1.1	11.4±43

Absobse cell numbers were calculate absobse lycaphocyte cell course.

Lymphocyte Subsets (Absolute Numbers) in Rat Thymus Absolute Cell Number (X107)/Thymus (MEAN ± SD)

FEMALE

LYMPHOCYTE SUBSETS	CONTROL	PAIR-FED CONTROL (p=4)	300 tog/kg/d (n=3)	600 mg/kg/d (n=3)	1200 103/105/d (v=5)
Total Leukocytes	14.1 ± 11.0	163±133	122 ± 1.7	30.1 ± 6.9	15.6 ± 10.2
CD43*	133±105	17.6±14.1	11.6 ± 1.6	28.5 ± 6.5	14.7 ± 9.8
CD3	11.4±9.1	129±102	9.7 ± 1.0	20.5 ± 6.5	125±83
CD3-CD4	11.2 ± 8.9	12.5 ± 9.9	9.5 ± 1.0	20.1 ± 6.4	12.2 ± 8.1
CD3.CD8.	10.6 ± 8.1	120±93	92109	19.9 ± 5.9	21.5 ± 7.7
IgM	0.7 ± 0.3	0.9±0.6	0.5 ± 0.2	1.2±0.6	0.6 ± 0.4
CD4°CD8-	0.6±0.4	0.8 ± 0.5	0.6±0.2	1.4±0.5	1.0 ± 0.6
CD4CD8*	Q.7±0.5	0.8 ± 0.6	0.5 ± 0.2	1.3 ± 0.3	0.9 ± 0.6
CD4*CD8+	12.6 ± 9.9	14.2 ± 11.9	10.9 ± 1.3	25.7 ± 5.9	13.4±9.2
CD4CD\$	Q.I ±Q.I	0.7 ± 0.1	0.2 ± 0.1	0.3±0.2	0.1 ± 0.1

Absolute cell numbers were calculated by multiplying the "corrected" lymphocyte subset percentages by the absolute lymphocyte cell count.

Abbreviations and Symbols:
n = The number of animals included in analysis

Toxicokinetics:

Dose (mg/kg/d)	- 300 = 100	mg/kg TID	600 = 200	mg/kg TID	1200 = 400 mg/kg TID	
Sex	. M	F	M	F	M	F
Day 1 C _{max} (μg/ml)	7.78	6.91	11.1	9.07	14.4	12.8
AUC _{0-8 hr} (μg.h/ml)	15.6	14.6	33.7	30.0	41.4	47.0
T _{max} (h)	1.0	1.0	1.0	1.0	1.0	1.0
Week 13 C _{max} (µg/ml)	9.65	9.42	11.1	22.3	17.4	34.0
AUC _{0-8 hr} (µg.h/ml)	22.3	22.9	39.8	46.3	59.9	72.1
T _{max} (h)	• 1.0	1.0	1.0	1.0	1.0	1.0
Week 26 C _{max} (µg/ml)	12.4	10.2	18.2	25.2	22.4	26.7
AUC _{0-8 hr} (µg.h/ml)	30.7	26.1	55.9	40.2	68.0	62.7
T _{max} (h)	1.0	0.5	1.0	1.0	1.0	1.0

Summary of Study Findings:

In this rat study, animals were dosed orally by gavage with OGT 924 at 300, 600 and 1200 mg/kg/d (administered as three equally divided doses at 8 hr intervals) for 26 week followed by a 4-week recovery period. Dose-related increase in scaly tail was observed in both sexes. Wartlike lesions on the tail also showed a dose-dependent increase in treated males. Mean body weight was decreased by 12% in MD males (6x the maximum clinical dose of 100 mg TID based on AUC_{0-6hr}) and by 23% and 9% in HD males (8x the maximum clinical dose of 100 mg TID based on AUC_{0-6hr}) and females (7x the maximum clinical dose of 100 mg TID based on AUC_{0-6hr}) respectively at the end of the treatment period. After the 4-week recovery period, mean body weight was still decreased by 10% in MD males and by 17% and 8% % in HD males and females respectively. Animals in the pair-fed control group also had decreased mean body weight (8%-10%) which was partially reversed to 6% at the end of the recovery period. Mean food consumption was decreased by 8% and 13% in MD and HD males and by 15% and 7% in the pair-fed control males and females respectively. At the end of the recovery period, mean food consumption showed partial recovery.

Sperm motility, sperm concentration and the number of normal sperms were decreased in all treated males relative to control. Amorphous sperms and sperms with decapitated heads were increased in all treated males relative to control. At the end of the treatment-free period, partial recovery of the altered sperm parameters was observed. It is likely full recovery will occur with time.

The target organs of toxicity include the GI tract – esophagus (serosal fibrosis), stomach (cytoplasmic vacuolation of chief cells), large intestine (mucosal necrosis), pancreas (acinal cell vacuolation), salivary gland (cytokaryomegaly – seromucus acinar cells), epididymides (hypospermia), testes (degeneration/atrophy), mesenteric lymph node (lymphoid depletion), skin of the tail (hyperkeratosis, acanthosis, pyogranulomatous dermatitis), eye (posterior synechia, cataract) and kidney (chronic progressive nephropathy). The incidence of testicular lesions was decreased at the end of the recovery period but the severity appear to have increased. The incidence of kidney lesions was decreased at the end of the recovery period. NOAEL could not be established because of the nephropathy and testicular lesions at the LD (3x the maximum clinical dose of 100 mg TID based on AUC_{0-6hr}).

STUDY TITLE: One-year oral toxicity study of SC-48334 in rats with a 4-week recovery period.

Key study findings:

 7/60, 22/60, 23/60 and 18/60 animals were observed with palpable masses in the purina control, LD, MD and MHD groups, respectively. No dose-relationship was apparent in either the total number of animals affected (or the onset of occurrence at these dose levels. Only 2/60 animals in the HD group had palpable masses, however, this group was sacrificed early (during study week 20). In general, the masses were observed during the last month of treatment; masses were only observed in 3/60, 5/60, 12/60 and 6/60 animals in the control, LD, MD and MHD groups, respectively, during the last four weeks of dosing. Macroscopic and/or microscopic examination of the masses revealed them to be abscesses and/or associated with the mammary glands (lactation, galactoceles or active secretion). Only 3 masses were found to be tumors; an adenoma was present in 1/60 LD animal and 2/60 control group animals each had a fibroadenoma.

- Mortality occurred at all dose levels including controls. Due to the high mortality at HD, dosing was terminated during week 10 and sacrificed during week 20, 44/60 (HD), 15/60 (HMD), 9/60 (MH), 8/60 (LD), 3/60 (diet control) and 11/60 (— diet control) animals were found dead or sacrificed moribund during SC-48334 administration. The cause of death or moribundity was not ascertained in 35/60 (HD), 7/60 (HMD), 2/60 (MD), 6/60 (LD), 3/60 (purified diet control) and 7/60 (— diet control) animals. The causes of death established by sponsor include septicemia (1/60-HD, 1/60-MD); chronic renal disease (1/60-HD, 3/60-HMD, 2/60-MD); enteropathy (5/60-HD); gavage error (1/60-HD, 2/60-each for HMD, MD & LD); trauma (1/60-HD, 2/60-MD); pituitary tumor (1/60-HMD, 2/60-MD) and leukemia (1/60- — diet control). Reviewer believes that demise of most animals was due to GI toxicity (stomach-ulcer, hyperkeratosis; cecum-mucosal necrosis, inflammation, hemorrhage colon-dilatation of crypts, necrosis, edema, inflammation, ileum and jejunumvillous atrophy), renal toxicity (nephrosis, nephropathy, vacuolation of tubular epithelium, inflammation, protein casts and mineralization of tubular epithelium and pelvis), hepatotoxicity (necrosis, cytoplasmic vacuolation, hemorrhage and lymphocytic infiltrate) and cardiac toxicity (cardiomyopathy, inflammation and necrosis of myocardial fiber) based on the histopathology data provided.
- A dose-dependent decrease in % mean body weight was observed in both treated males and females. At 420 mg/kg/d, the % decreases in mean body weight were 18% and 14% respectively for males and females. At 840 mg/kg/d, the values were 32% and 26% for males and females respectively.
- Diarrhea was observed at doses ≥ HMD. The incidence of diarrhea was very high especially in the HD group during the first 12 weeks of treatment (45/60 weeks 1-4; 46/60 weeks 5-8; 21/60 weeks 9-12). Treatment was terminated in the HD group at week 10 due to mortality. However, the diarrhea persisted till week 12 and thereafter subsided completely. In the HMD animals, the incidence of diarrhea was 26/60 during the first four weeks of treatment but subsided significantly from week 5 onwards.
- Percent decrease in food consumption also showed a dose-dependent effect. At 420 mg/kg/d, the % decreases in food consumption were 13% and 5% respectively for males and females. At 840 mg/kg/d, the values were 26% and 9% for males and females respectively.
- Equatorial cataracts were observed in treated males relative to controls. The incidence appears to be dose-related. In treated males the incidence are1/28 (180mg/kg), 1/29 (420 mg/kg) and 18/27 (840 mg/kg) at week 52. In treated females, the incidence at the 840 mg/kg dose is 9/23 at week 52. After the 4 week recovery period, the incidence had decreased to are1/10 (180mg/kg), 1/9 (420 mg/kg) and 5/9 (840 mg/kg) in treated males and 4/8 for treated females. In the 1680 mg/kg dose group that was terminated at week 20, the incidence of cataracts was 9/9 (males) and 5/7 (females) at week 14.
- WBC increased in a dose-dependent manner achieving statistical significance in all treated males and in females dosed 420 and 840 mg/kg/d. RBC was slightly but statistically significantly decreased in males dosed 180 and 420 mg/kg/d and in females dosed 420 and 840 mg/kg/d. MCH was significantly increased in males dosed 420 mg/kg/d but decreased in females dosed 180 mg/kg/d. While MCHC was slightly but statistically significantly

decreased in all treated females, it was only decreased in males dosed 840 mg/kg/d. Platelets were dose-dependently and statistically significantly decreased in 420 and 840 mg/kg/d males and in the 840 mg/kg/d females. At the end of the recovery period, all affected hematology parameters had returned to normal limits.

- At week 52, K, P, BUN and Ca were slightly but statistically significantly increased in all treated males relative to control. K was slightly but statistically significantly increased in all treated females. P was dose-dependently and slightly but statistically significantly increased in females dosed 420 and 840 mg/kg/d. Creatinine and cholesterol levels were also significantly increased in males dosed 180 and 420 mg/kg/d. Mg was slightly but statistically significantly increased in the 180 mg/kg/d males. AST and ALT increased in a dose-dependent manner achieving statistical significance in males dosed 420 and 840 mg/kg/d and in females dosed 840 mg/kg/d. At the end of the recovery period, P and Na were slightly but statistically significantly increased in males and females dosed 840 mg/kg/d respectively.
- At the end of the treatment period (week 52), urine Ca was dose-dependently and statistically significantly increased in all treated males and in females dosed 420 and 840 mg/kg/d. Urine volume was statistically significantly increased in all treated females. Urine total protein was statistically significantly increased in males dosed 180 and 420 mg/kg/d but not in the 840 mg/kg/d group. All urinalysis parameters returned to normal limits at the end of the recovery period.
- Relative weight of the liver was slightly but statistically significantly increased in the 180 and 420 mg/kg/d males and in females dosed 420 and 840 mg/kg/d. Since there is no correlative histopathology, the increment may be due to the increased AST and ALT levels. The weight changes in the liver were partially reversed in females and fully reversed in males at the end of the recovery period. Absolute weight of the epididymides decreased in a dose dependent manner achieving statistical significance in at the 420 and 840 mg/kg/d dose levels. At the end of the recovery period the 840 mg/kg/d males still had a significantly decreased epididymal weight. Weights (abs. and rel.) of the testes decreased in a dose-dependent manner achieving statistical significance at the 840 mg/kg/d dose level. The decreased weights persisted throughout recovery. The decreased relative weight of the testes correlates with the atrophy of the seminiferous tubules. Relative weight of the kidney was statistically significantly increased in females dosed 420 and 840 mg/kg/d. At the end of the recovery period, the relative kidney weight of females dosed 420 mg/kg/d was still significantly different from control. In males, while weights were not significantly different from controls after treatment, relative kidney weight was statistically significantly increased in males dosed 180 and 840 mg/kg/d at the end of the recovery period. This may be due to the hyperplasia, nephropathy and mineralization of the pelvis observed histologically at the end of the recovery period.
- The target organs of toxicity include the epididymides (hypospermia), kidney (nephropathy, lymphocyte infiltration, protein casts, hyperplasia and mineralization), GI tract (stomachulcer, hyperkeratosis; cecum-mucosal necrosis, inflammation, hemorrhage colon-dilatation of crypts, necrosis, edema, inflammation; ileum and jejunum-villous atrophy), testes (atrophy of seminiferous tubules, aspermatogenesis, edema, and hyperplasia of interstitial cells), mammary gland (galactocele, active secretion) and tail (suppurative inflammation). Most of these toxicities occurred at all dose levels and showed very little/no recovery at the end of the treatment free period.
- NOAEL could not be established since toxicities were observed at the LD tested.

Study no: PSA-91C-3490

Volume #, and page #: Vol. 3; pg. 955.

Conducting laboratory and location:

Date of study initiation: May 1989. GLP compliance: Yes (USA, Japan).

QA report: yes (x) no ()

Drug, lot #, radiolabel, and % purity: Lot #s 88K040-301I, 88K042-301I, 89K008-301J. No

information on purity.

Formulation/vehicle: SC-48334 dissolved in distilled/deionized water.

Methods (unique aspects):

Dosing: Animals were dosed orally (gavage) with the test article at 180, 420, 840 and 1680 mg/kg/day divided into three equal doses administered TID at intervals of 8 hr apart. All dose groups received a diet of _____ rodent chow except the Control group 2 and the 1680 mg/kg/d group that received purified diet (dextrose). The purified diet was used to minimize the test article-related diarrhea observed at higher dose levels in previous studies with SC-48334.

Species/strain: Rat/Crl:CD.BR

#/sex/group or time point (main study): 30/sex/group.

Satellite groups used for toxicokinetics or recovery: 20/sex/group used for TK.

Age: 5 weeks at study initiation.

Weight: 96 - 175 g (M); 90 - 148 g (F).

Doses in administered units: 180, 420, 840 and 1680 mg/kg/day.

Route, form, volume, and infusion rate: Oral (gavage), solution, 10 ml/kg.

Observations and times:

Clinical signs: Daily.
Body weights: Weekly.
Food consumption: Weekly.

Ophthalmoscopy: Conducted prior to study initiation on all animals and during weeks 9, 14, 26,

50 and 55.

EKG: Not evaluated.

Hematology: Blood samples were collected from animals fasted overnight. Blood samples were taken from 10 male and 10 female rats in control group 2 (purified diet) and all rats in the 1680 mg/kg/day toxicology group after 10 weeks of test article administration (reticulocyte and differential white blood cell counts only) and at the interim sacrifice (week 20), from 10 male and 10 female rats from each toxicology study group after approximately 16 and 32 weeks of test article administration, and from all animals at the end of dosing (week 52) and at the end of the recovery period (week 56).

Clinical chemistry: Blood samples were collected as indicated for hematology. Routine clinical chemistry was conducted.

Urinalysis: Urine samples were collected from the animals bled weeks 10, 16, 20, 32, 52 and 56. A 24-hour urine sample was collected from each animal using a metabolism cage. Urine samples were generally collected approximately one week prior to blood samples.

Gross pathology: Organs/tissues collected for gross pathology examination is indicated in the list of addendum.

Organs weighed: Organs weighed at the interim (week 20), terminal (week 52) and recovery (week 56) sacrifices are indicated in the list of addendum.

testes and thymus from all animals in the 180 and 420 mg/kg/day toxicology groups at the week 52 sacrifice and all animals at the recovery (week 56) sacrifice were examined histologically. Toxicokinetics: Blood samples for PK evaluation were collected from animals in the PK groups on the first day of dosing and during study weeks 10, 25 and 51 (weeks 11, 26 and 52 of dosing, respectively). On the first day of dosing, blood samples were collected from all animals (20/sex/group) at appropriate intervals at approximately 0.5, 1, 2, 4 and 8 hours after dosing). Due to the high mortality observed in the HD (1680 mg/kg/d) group, dosing was terminated during week 10 and all animals were sacrificed. Prior to sacrifice and following the last dose, blood samples were collected from all HD animals. Blood samples were collected during weeks 25 and 51 (week 26 and 51 of dosing) from 15 animals/sex in the 180, 420 and 840 mg/kg/day groups.

Results:

Мо	72	11817
1710	110	HLΥ.

Dose (mg/kg/d)	0	0	180	420	840	1680
Males	diet)	(Purified diet)		Í		i
Found Dead	1/30 (3)		1/30 (37)	1/30 (29)	1/30 (35)	1/30 (0)
Found Dead	1/30 (11)		1/30 (44)		1/30 (43)	2/30 (3)
Found Dead	1/30 (15)				1/30 (44)	1/30 (4)
Found Dead	2/30 (26)				1/30 (51)	3/30 (6)
Found Dead						3/30 (7)
Found Dead						1/30 (8)
Sacrificed Moribund						3/30 (3)
Sacrificed Moribund						3/30 (4)
Sacrificed Moribund						2/30 (7)
Sacrificed Moribund			1			1/30 (9)
Sacrificed Moribund]		1/30 (10)
Total Deaths	5/30	0/30	2/30	2/30	5/30	21/30*
Scheduled Sacrificed	15/25 (52)	30/30 (20)	18/28 (52)	19/29 (52)	16/26 (52)	9/9 (20)
Found Dead				1/10 (54)	1/10 (53)	
Recovery Sacrifice	10/10 (56)		10/10 (56)	9/9 (56)	9/9 (56)	
Dose (mg/kg/d)	0	0	180	420	840	1680
Females	(diet)	(Purified diet)	}		1	
Found Dead	1/30 (4)	1/30 (2)	1/30 (1)	1/30 (15)	1/30 (0)	2/30 (2)
Found Dead	2/30 (15)	2/30 (106)	1/30 (14)	1/30 (22)	2/30 (15)	2/30 (4)
Found Dead	1/30 (17)		3/30 (15)	1/30 (47)	1/30 (20)	1/30 (5)
Found Dead	1/30 (45)				1/30 (27)	1/30 (6)
Found Dead					1/30 (31)	1/30 (8)
Found Dead					2/30 (50)	3/30 (9)
Found Dead					2/30 (52)	1/30 (10)
Sacrificed Moribund	1/30 (135)		1/30 (39)	1/30 (38)		1/30 (3)
Sacrificed Moribund				1/30 (40)		3/30 (4)
Sacrificed Moribund				1/30 (46)	<u> </u>	4/30 (5)
Sacrificed Moribund				1/30 (51)		2/30 (6)
Sacrificed Moribund						2/30 (7)
Total Deaths	6/30	3/30	6/30	7/30	10/30	23/30°
Interim sacrifice						
Scheduled Sacrificed	15/25 (52)	28/28 (20)	15/24 (52)	15/23 (52)	12/20 (52)	7/7 (20)
Recovery Sacrifice	9/9 (56)		9/9 (56)	8/8 (56)	8/8 (56)	
M + F Deaths	11/60	3/60	8/60	9/60	15/60	44/60

^{*} Dosing stopped at week 10; Number in parenthesis indicate week of occurrence.

Histopathology:

		SU	GGESTE	D CAU	SE OF	DEATH	(BY SP	ONSOR)				
G	Pross P	atholo	gy of A	Animals	s Four	d Dea	d or Sa	crifice	d Morit	ound		
DOSE (mg/kg/d)		diet	Purific	0 ed diet	1	В0	4	20	8	40	16	80
Sex	M	F	M	F	M	F	M	F	M	F	M	F
Animals/group	30	30	30	30	30	30	30	30	30	30	30	30
Animals found dead/sacrificed	5	. 6	0	2	2	6	2	7	5	10	21	23
Intubation error	2/5	3/6			1/2		1/2	1/7		2/10	1/21	1/23
Atrial thrombus							1/2			l'		
Died during sinus blood collection										2/10		
Chronic renal dzs.								2/7	2/5	1/10		1/23
Septicemia								1/7				
Pituitary tumor			1					2/7		1/10		
Enteropathy												2/23
Mech. Trauma												_
Vertebral column									<u> </u>	L		1/23
Cause of death												
Undetermined	3/5	3/6	<u> </u>	2/2	1/2	6/6		1/7	3/5	4/10	20/21	18/23

Clinical signs: Numerous clinical signs of toxicity were observed in the HD animals. The predominant signs consisted of atonia, unkempt appearance, hypothermia, red and swollen forepaws, a reddened distal end of the tail, redness of the anal orifice, a swollen lower lip of the mouth, puffiness of the extremities and facial area, exfoliation of the tail (medial to distal), apparent segmenting of the tail, scabbing on the nose, red, brown or yellow material and/or staining on the anogenital, urogenital and diarrhea. Although these findings were generally observed at more than one observation period, the majority of these findings were most prevalent one hour following the first daily dose. In addition, several findings (redness of the anal orifice, diarrhea and feces with red material) were generally observed more frequently in females than in males. Most of these findings were not observed following the termination of dosing (during week 10) or the frequency greatly decreased during the recovery period.

Similar findings judged to be related to treatment were noted at the lower dose levels of LD, MD and MHD. This included the following: the distal end of the tail was reddened in MD and MHD animals; the incidence of this finding was markedly decreased during the final three to four months of dosing. Exfoliation of the tail (medial to distal) and apparent segmenting of the tail occurred at the LD, MD and MHD levels. The frequency and onset of occurrence were doserelated, and the incidence of both findings generally decreased during the 4-week recovery period. The only other finding that appeared to be related to test article administration at the LD, MD and MHD was the apparent raised areas on the mid and proximal tail. The frequency of this finding was dose-related and greatly decreased during the recovery period.

Palpable ventral masses were noted in males and females in control group 1 and in the LD, MD, MHD and HD groups. During the study, more animals in the LD, MD and MHD groups had masses than in the control group. No dose-relationship was apparent in either the total number of animals affected (7/60, 22/60, 23/60 and 18/60 animals with masses in the control, LD, MD and MHD groups, respectively) or the onset of occurrence at these dose levels. Only 2/60 animals in the HD group had palpable masses, however, this group was sacrificed early (during study week 20). Masses were only observed in 3/60, 5/60, 12/60 and 6/60 animals in the control, LD, MD and MHD groups, respectively, during the last four weeks of dosing. Macroscopic and/or microscopic examination of the masses revealed them to be abscesses and/or associated with the mammary glands (lactation, galactoceles or active secretion). Only 3

masses were found to be tumors; an adenoma was present in 1/60 LD animal and 2/60 control group animals each had a fibroadenoma.

Incidence of Diarrhea: empty cells indicate zero incidence.

DOSE (mg/kg/d)	0 — diet	0 Purified diet	180	420	840	1680
Weeks 1 - 4					26/60	45/60
Weeks 5 - 8		1			0/60	46/60
Weeks 9 - 12					0/60	21/60
Weeks 13-16					2/60	0/60
Weeks 17- 52					1/60	•
Weeks 53-56 (Recovery)					0/60	1

^{*} Treatment was terminated at week 10, sacrificed at week 20

Body weights: Males (g)

DOSE (mg/kg/d)	0 diet	0 Purified diet	180	420	840	1680
Pretest N	142 ± 15.4 30	141 ± 12.0 30	136 ± 20.4 30	136 ± 13.3 30	136 ± 13.6 30	140 ± 12.6 30
Week 20 N	544 ± 61.9 26	547 ± 58.1 30	521 ± 53.6 30	406 ± 40.8** 29	412 ± 68.1** 30	476 ± 27** 9
Week 52 N	663 ± 86.4 25	Α	604 ± 74.8* 28	545 ± 44.2** 29	454 ± 67.1** 26	Α
Week 56 Recovery: N	669 ± 44.3 10	Α	592 ± 63.3* 10	577 ± 52.6* 9	459 ± 67.2** 9	Α
% ↓ Mean B. wt.	0		9	18	32	

A = interim sacrifice at week 20; * p < 0.05; ** p < 0.01

Females (g)

DOSE (mg/kg/d)	n —— jiet	0 Purified diet	180	420	840	1680
Pretest N	129 ± 8.1 30	126 ± 8.0 30	124 ± 8.0 30	124 ± 9.9 30	125 ± 7.4 30	124 ± 9.3 30
Week 20 N	366 ± 48.6 25	348 ± 48.6 28	352 ± 44.7 25	354 ± 40.9 29	312 ± 32.9** 27	320 ± 29 7
Week 52 N	468 ± 92.2 24	Α	429 ± 66.9 24	402 ± 40.5** 23	348 ± 34.7** 26	Α
Week 56 Recovery: N	479 ± 83.3 9	Α	438 ± 68.9 9	407 ± 53.3 8	409 ± 40.7 8	Α
% ↓ Mean B. wt.	0		8	14	26	

[%]Mean B. wt. at week 52; A = interim sacrifice at week 20; * p < 0.05; ** p < 0.01

Food consumption: Males (g/animal/day)

DOSE (mg/kg/d)	0 diet	0 Purified diet	180	420	840	1680
Pretest N	21 ± 2.3 30	19 ± 2.0** 30	20 ± 2.2 30	19.8 ± 1.7 30	19 ± 2.1** 30	16 ± 4.9** 29
Week 20 N	27 ± 2.9 26	27 ± 3.3 30	28 ± 3.1 30	24 ± 2.7 29	25 ± 3.7 30	27 ± 3.3 9
Week 52 N	23 ± 4.3 25	A	21 ± 2.9 28	20 ± 4.1 29	17 ± 4.61** 26	Α
Week 56 Recovery: N	24 ± 3.5 10	A	24 ± 2.5 10	24 ± 2.7 9	22 ± 4.2 9	A
% ↓ Fd. Con.	0		9	13	26	· _

% \downarrow Fd. Con. At week 52; A = interim sacrifice at week 20; *p < 0.05; **p < 0.01

Females (g/animal/day)

DOSE (mg/kg/d)	0 diet	0 Purified diet	180	420	840	1680
Pretest N	20 ± 1.7 30	20 ± 3.0 29	19 ± 2.2 30	19 ± 1.8 30	18 ± 1.9 29	16 ± 4.6** 30
Week 20 N	22 ± 3.2 25	25 ± 4.4* 28	23 ± 3.1 25	25 ± 3.2* 29	21 ± 2.8 27	27 ± 6.1° 7
Week 52 N	22 ± 3.5 24	A	22 ± 2.7 24	21 ± 2.7 23	20 ± 2.7 22	Α
Week 56 Recovery: N	22 ± 3.8 9	Α	22 ± 3.3 9	22 ± 2.6 8	24 ± 3.7 8	A
% ↓ Fd. Con.	0		0	5	9	

% \downarrow Fd. Con. At week 52; A = interim sacrifice at week 20; *p < 0.05; **p < 0.01

Ophthalmoscopy:

Males (equatorial cataracts/animals examined)

DOSE (mg/kg/d)	n	0	180	420	840	1680
	diet	Purified diet		ļ		
Week 9	0/29	0/30	1/30	1/30	16/30	7/10
Week 14	0/28	0/30	0/30	1/30*	18/30	9/9
Week 26	0/26		0/30	2/30°	24/30	
Week 52	1/25		1/28*	1/29**	18/27	,
Week 55:recovery	0/10		1/10*	1/9**	5/9	-

^{*} Or ** = same animal

Females (equatorial cataracts/animals examined)

DOSE (mg/kg/d)	0	0	180	420	840	1680
	diet	Purified diet		į.	,	1
Week 9	0/29	0/29	0/29	0/30	1/29	2/8
Week 14	0/29	0/29	0/29	3/30	12/28	5/7
Week 26	0/25		0/25	3/28	14/26	
Week 52	0/24		0/24	0/24	9/23	
Week 55:recovery	0/9		0/9	0/8	4/8	

Electrocardiography: No data.

Hematology:

Hematology.				· · · · · · · · · · · · · · · · · · ·		
		Males -	Week 52 data)		
DOSE (mg/kg/d)	0	0	180	420	840	1680
	diet	Purified diet		İ		
WBC (10 ³ /UL)	10.6 ± 2.3	Α	16.0 ± 4.9°	17.9 ± 6.2**	19.3 ± 9.0**	Α
N	24	<u> </u>	26	28	25	
RBC (10 ⁵ /UL)	8.5 ± 0.4	Α	8.0 ± 0.7°	7.9 ± 0.8**	8.3 ± 0.7	Α
MCV (U3)	55.0 ± 2.1	Α	56.6 ± 2.2	57.5 ± 2.8**	56.7 ± 3.01	Α
MCHC (%)	34.7 ± 0.8	Α	34.6 ± 1.1	34.3 ± 1.1	33.5 ± 1.2**	Α
PLT (10 ³ /UL)	1123 ± 188	Α	1146 ± 182	852 ± 162**	782 ± 212**	Ä
		Females -	- Week 52 dat	la		
DOSE (mg/kg/d)	0	0	180	420	840	1680
(0 0 ,	diet	Purified diet			'	
WBC (103/UL)	8.0 ± 3.0	Α	10.0 ± 2.1	11.9 ± 3.5**	13.0 ± 3.1**	Α
N	23	ł	22	22	22	
HGB (g/dl)	15.6 ± 1.0	Α	15.2 ± 0.68	14.7 ± 0.6**	15.0 ± 0.6°	A
MCH (pg)	21.6 ± 1.0	Α	20.8 ± 0.8*	21.1 ± 1.1	20.5 ± 0.93	Α
MCHC (%)	36.8 ± 1.36	Α	35.0 ± 0.75**	35.2 ± 1.1**	34.3 ± 1.2**	Α
PLT (103/UL)	1023 ± 118	Α	1090 ± 136	933 ± 180	828 ± 137**	Α

A = interim sacrifice at week 20; * p < 0.05; ** p < 0.01

By the end of the recovery period (week 56), all affected hematology parameters had returned to normal limits.

Clinical chemistry:

		Males - 1	Week 16 data			
DOSE (mg/kg/d)	n	0	180	420	840	1680
	diet	Purified diet				
AST (U/L)	125 ± 10.3	121 ± 15.1	141 ± 40.3	195 ± 90.7°	299 ± 99.8**	128 ± 25.7
Phosphorus (mg/dl)	9.1 ± 1.1	8.0 ± 1.4	10.1 ± 1.5	11.6 ± 2.5**	10.4 ± 1.3	8.2 ± 1.5
T Bilirubin (mg/dl)	0.2 ± 0.05	0.2 ± 0.05	0.2 ± 0.05	0.3 ± 0.05	0.3 ± 0.08**	0.3 ± 0.1
	•	Males - 1	Week 52 data	1		
AST (U/L)	156 ± 51.6	Α	158 ± 56.9	227 ± 98.1**	352 ± 89.7**	Α
ALT (U/L)	74 ± 23.6	Α	69 ± 17.2	106 ± 44.1**	137 ± 37.5**	Α
ALKP (U/L)	82 ± 18.4	Α	46 ± 15.2**	65 ± 22.2**	62 ± 18.1**	Α
Phosphorus (mg/dl)	7.2 ± 1.0	Α	9.1 ± 2.6°	9.2 ± 2.8**	9.9 ± 2.1**	Α
BUN (mg/dl)	12.2 ± 1.7	Α	15.0 ± 4.0*	16.3 ± 4.6**	15.6 ± 2.1**	Α
Creatinine (mg/dl)	0.6 ± 0.07	Α	0.8 ± 0.15**	0.7 ± 0.14**	0.7 ± 0.10	_ A
Cholesterol (mg/dl)	74 ± 21.7	Α	110 ± 43**	107 ± 44°	91 ± 42.3	Α
Calcium (mg/dl)	11.3 ± 0.88	Α	12.1 ± 0.90*	12.1 ± 0.97**	12.3 ± 0.93**	Α
Potassium (mEq/l)	5.91 ± 0.6	Α	6.94 ± 0.7**	6.87 ± 0.93**	7.52 ± 1.1**	Α
Males - Recover	y Week 56					
Phosphorus (mg/dl)	7.2 ± 0.65	Α .	7.9 ± 1.14	8.2 ± 1.02	8.5 ± 0.68*	Α
		Females ·	Week 16 dat	ta	·	; ,
DOSE (mg/kg/d)	0	0	180	420	840	1680
	diet	Purified diet				
AST (U/L)	130 ± 19.1	107 ± 16.6	138 ± 18.2	143 ± 18.7	251 ± 60.3**	102 ± 12.6
ALT (U/L)	83 ± 21.1	49 ± 8.9**	59 ± 5.7**	58 ± 9.5**	78 ± 12.1	45 ± 5.4**
T Protein	7.5 ± 0.9	7.0 ± 0.4	7.1 ± 0.6	7.1 ± 0.6	6.5 ± 0.5**	6.6 ± 6.3*
Albumin	4.7 ± 0.5	4.2 ± 0.3	4.5 ± 0.5	4.3 ± 0.4	3.9 ± 0.5**	3.9 ± 0.3**
		Females -	Week 52 dat	ta		
AST (U/L)	149 ± 39.8	Α	133 ± 36.7	145 ± 24.3	265 ± 148**	Α
Phosphorus (mg/dl)	6.9 ± 1.2	Α	7.9 ± 1.7	8.1 ± 1.3*	8.4 ± 1.5**	Α
Potassium (mEq/l)	5.21 ± 0.62	Α	5.93 ± 0.93*	5.85 ± 0.82*	6.04 ± 0.98**	Α
Females - Recovery						
Sodium (mEq/l)	143 ± 1.2	A	144 ± 1.5	144 ± 1.2	145 ± 1.8*	Α

A = interim sacrifice at week 20; * p < 0.05; ** p < 0.01

Urinalysis:

		Males -	Week 15 data	1		
DOSE (mg/kg/d)	0 diet	0 Purified diet	180	420	840	1680
T. Volume (ml)	24.1 ± 6.3	16.8 ± 8.5	37.8 ± 14.1**	38.3 ± 7.01**	38.7 ± 9.3**	15.4 ± 4.9
Urine Ca (mg/dl)	5.1 ± 1.7	3.3 ± 1.62	9.4 ± 5.1	18.1 ± 9.4**	27.2 ± 10.3**	3.9 ± 1.11
		Males -	Week 51 data	1		
Urine Ca (mg/dl)	5.2 ± 2.42	Α	13.3 ± 5.40**	19.7 ± 6.24**	23.7 ± 7.48**	Α
Urine T. protein (mg/dl)	148 ± 160	Α	656 ± 515**	438 ± 346*	285 ± 337	Α
Males - Recovery W	leek 56					
	All a	iffected paramete	ers returned to no	rmal limits.		
		Females -	- Week 15 da	ta		
DOSE (mg/kg/d)	0	0	180	420	840	1680
	diet	Purified diet		<u> </u>		
T. Volume (ml)	21.3 ± 5.12	19.2 ± 10.0	30.7 ± 11.3	32.3 ± 16.0	36.4 ± 11.4*	20.4 ± 7.3
Urine Ca (mg/dl)	9.9 ± 4.0	4.1 ± 3.3	22.5 ± 9.24**	29.7 ± 8.27**	32.7 ± 8.14**	5.0 ± 2.22
Urine CI (mEq/I)	110 ± 57.1	29 ± 13.1**	76 ± 29.1	70 ± 20.3*	78 ± 25.3	36 ± 10.3**
Urine Na (mEq/l)	56 ± 16.4	29 ± 9.5**	43 ± 17.1	39 ± 8.9*	35 ± 15.7°	27 ± 9.0**
Urine K (mEq/l)	155 ± 48.7	66 ± 26.2**	109 ± 41.1*	103 ± 32.5**	93 ± 22.8**	67 ± 15.0**
		Females -	- Week 52 dat	la		
T. Volume (ml)	34.2 ± 10.6	Α	46.3 ± 14.3°	53.0 ± 20.9**	48.8 ± 15.3*	Α
Urine Ca (mg/dl)	17.0 ± 5.9	Α	21.3 ± 7.5	26.7 ± 10.4**	28.9 ± 8.9**	Α

Females - Recovery Week-56

All affected parameters returned to normal limits.

A = interim sacrifice at week 20; * p < 0.05; ** p < 0.01

Organ weights: Absolute organ weights. Only animals in the control (purified diet) and HD groups were sacrificed at week 20. Relative weights = organ wt. relative to final body wt. (g/100

g)		14	-1	M/2-1/ 20	-1-4				
D0051 " / "			ies -	Week 20	oat			_	4000
DOSE (mg/kg/d)	0	0	هم:ام	180		420	840	1	1680
	diet	Purified	a aiet	i		, i		1	
Kidney (g)	- Giet	3.44 ±	0.36	 			<u> </u>	20	± 0.18**
Liver (g)		15.8 ±							4 ± 0.87**
Epididymides (g)		1.41 ±							9 ± 0.10**
Epididymides (g/100 g	, 	0.27 ± 0						+	
Testes (g)	"	3.3 ± 0					 		± 0.027**
Testes (g/100 g))		0.62 ±							£ ± 0.33**
Brain (g/100 g)		0.41 ±							6 ± 0.08** 6 ± 0.03**
Spleen (g/100 g)	- - - 	0.15 ± 0							8 ± 0.26*
Pituitary gl. (g/100 g)		0.005 ±		 					6 ± 0.002°
Fituliary 91. (9/100 9)	!		_	Week 52	dat	a	L	1 0.00	0 T 0.002
Heart (g)	1.88 ± 0.19	A		7 ± 0.26		1.71 ± 0.26	1.53 ± 0.2	20**	A
Heart (g/100 G)	0.29 ± 0.04	A		0 ± 0.25	_	0.32 ± 0.04	0.34 ± 0.4		1 A ·
Liver (g)	19.8 ± 3.65	Â		9 ± 3.97		19.2 ± 3.12	16.1 ± 3.		A
Liver (g/100 g)	3.06 ± 0.32	Â	+	± 0.47**		3.62 ± 0.60**	3.47 ± 0		A
Salivary glands (g)	0.86 ± 0.11	A	4	7 ± 0.12		0.74 ± 0.11**	0.64 ± 0.0		A
Spleen (g)	0.80 ± 0.17	A		4 ± 0.16		0.83 ± 0.16	0.74 ± 0.0		A
Epididymides (g)	1.42 ± 0.14	A	+	7 ± 0.16		1.03 ± 0.19**	$0.74 \pm 0.$		A
Testes (g)	3.41 ± 0.37	Â		0 ± 0.76	_	2.90 ± 0.89	1.49 ± 0.4		A
Testes (g/100 g)	0.53 ± 0.07	A		6 ± 0.76	_	0.55 ± 0.16	0.33 ± 0.3		A
Brain (g/100 g)	0.35 ± 0.07	Â	+	8 ± 0.05		0.33 ± 0.10	0.33 ± 0.0		A
Adrenal gl. (g/100 g)									1 A
Pituitary gl. (g/100 g)	0.008 ± 0.0023 0.002 ± 0.0006	_		2 ± 0.0021		012 ± 0.0037** 003 ± 0.0005	0.012 ± 0.0 0.003 ± 0.0		1 Â
Males - Recovery We		<u>, , </u>	0.002	2 ± 0.0003	<u> </u>	003 ± 0.0003	1 0.003 ± 0.0	004	
Salivary glands (g)	0.80 ± 0.067	A	0.8	2 ± 0.07	-	0.75 ± 0.01°	0.66 ± 0.1	11**	A
Brain (g/100 g)	0.34 ± 0.03	A		4 ± 0.05		0.38 ± 0.03	0.47 ± 0.0		A
Kidneys (g)	4.06 ± 0.48	A		7 ± 0.50		3.89 ± 0.59	$3.35 \pm 0.$		A
Kidneys (g/100 g)	0.62 ± 0.05	A		2 ± 0.09°		0.69 ± 0.07	0.75 ± 0.0		A
Epididymides (g)	1.44 ± 0.17	A		0 ± 0.23		1.25 ± 0.24	0.83 ± 0.		A
Testes (g)	3.27 ± 0.94	A		1 ± 0.63		2.96 ± 0.97	1.11 ± 0.2		A
Testes (g/100 g)	0.50 ± 0.13	A		7 ± 0.14		0.53 ± 0.18	0.25 ± 0.0		A
Thyroid gland (g)	0.04 ± 0.004	A		3 ± 0.006	_	026 ± 0.005**	0.026 ± 0.0		
Heart (g/100 g)	0.27 ± 0.03	Α		9 ± 0.03		0.29 ± 0.02	0.37 ± 0.1		A
Adrenal gl. (g/100 g)	0.009 ± 0.002			0 ± 0.003		.010 ± 0.002	0.014 ± 0.0	_	A
Pituitary gl. (g/100 g)	0.003 ± 0.0008		·	± 0.0005		002 ± 0.0003	0.003 ± 0.0		A
	, 5.002 2 0.000			- Week 20				- 3 ·	
DOSE (mg/kg/d)	0	0 Purified	dica	180		420	840		1680
Kidney (g)	diet	2.67 ± (<u> </u>	1 2	2 ± 0.22**
Liver (g)		12.2 ± 2							93 ± 0.91**
Liver (g/100 g)		3.58 ± (-		98 ± 0.91
Liver (g/100 g)				- Week 52	2 da	ta			30 I U.24
Salivary glands (g)	0.66 ± 0.08	Α		0.62 ± 0.0		0.63 ± 0.10	0.56 ± 0.0	7**	Α
Ovaries (g)	0.12 ± 0.023	Α		0.11 ± 0.0		0.11 ± 0.019	0.09 ± 0.0		A
Brain (g/100 g)	0.46 ± 0.099	Α		0.49 ± 0.0		0.53 ± 0.066	0.61 ± 0.03		Α

Heart (g/100 g)	0.33 ± 0.055	A	0.35 ± 0.053	0.37 ± 0.036	0.39 ± 0.028**	Α
Kidney (g/100 g)	0.71 ± 0.11	Α	0.82 ± 0.22	0.92 ± 0.22**	0.90 ± 0.10°	Α
Liver (g/100 g)	3.35 ± 0.43	Α	3.83 ± 0.72	4.12 ± 0.73**	4.03 ± 0.48**	Α
Females - Recover	y Week 56					
Heart (g/100 g)	0.31 ± 0.04	A	0.36 ± 0.07	0.39 ± 0.04°	0.40 ± 0.05**	A
Liver (g/100 g)	3.29 ± 0.44	Ā	3.91 ± 0.54	4.35 ± 0.71**	3.54 ± 0.18	Α
Kidney (g/100 g)	0.72 ± 0.08	Α	0.91 ± 0.24	1.08 ± 0.29**	0.84 ± 0.17	. A
Uterus (g/100 g)	0.20 ± 0.07	Α	0.31 ± 0.07°	0.27 ± 0.04	0.30 ± 0.09**	Α

A = interim sacrifice at week 20; * p < 0.05; ** p < 0.01

Gross pathology: Animals found dead or sacrificed moribund at all dose groups were necropsied. Empty cells indicate zero incidence for a particular finding.

Gross Pathology of Animals Found Dead or Sacrificed Moribund DOSE (mg/kg/d) 180 420 1680 **Purified diet** diet M F M F М M F 30 30 30 Animals/group 30 30 30 30 30 30 30 30 30 2 7 6 Ō 2 10 21 23 Animals found 6 dead/sacrificed Adrenal glands Dark red 2/5 1/2 1/7 3/5 3/10 2/21 6/23 Adrenal glands Reddened 1/5 1/6 2/7 1/23 **GI Tract** 2/5 6/6 6/6 7/7 1/5 2/10 4/21 1/23 Dark red contents 2/2 Stomach Dark red foci 1/6 3/7 6/21 6/23 Esophagus 1/2 1/10 Hemorrhagic Esophagus 1/10 Rupture Esophagus Perforation 2/6 Heart 1/2 1/5 Cardiomegaly Heart Atrial thrombus 1/2 Kidney Dark red 2/5 1/5 1/10 2/21 2/23 Kidney 1/10 1/23 Reddened corticomedullary junction Kidney 2/7 1/5 Nephrosis Kidney 1/5 1/10 1/23 White foci Lungs White foamy 2/5 1/6 1/2 1/10 1/23 contents Lungs 1/10 2/21 Mottled Lungs 2/6 1/10 1/5 Dark red Lungs Clear fluid contents 1/21 Pituitary Reddened 3/6 1/7 1/10 2/21 3/23 Pituitary 2/7 3/6 1/10 Enlarged Testes

Small 5/5

Only animals in the control (purified diet) and HD groups were sacrificed at Week 20.

DOSE (mg/kg/d)		n		nals Sac		80		20		40	46	80
DOSE (mg/kg/d)		diet		ed diet				LU		+0	"	000
Sex	M	F	M	F	M	F	М	F	М	F	M	F
Surviving												
Animals/group	26	25	30	28	30	25	29	29	30	26	9	7
# of Animals sacrif												
-iced at Week 20		<u>.</u>	30	28			Ĺ				9 `_	7
Lymph node, Sub.												
Reddened			4/30	6/28				i	j	ł	4/9	3/7
Lymph node, Sub.												
Enlarged			5/30	1/28						L	<u> </u>	
Kidney												
Rough				3/28				<u> </u>				1/7
Pituitary				•								
Enlatged				3/28				<u> </u>	<u> </u>			
Salivary glands		[
Hemorrhagic area									İ		1/9	_ `
Spinal cord		Ĭ	1								1	
Red fluid beneath		ì ·			i			1	Ì]	1/9	',
meninges			<u> </u>									<u> </u>
Testes												
Small			1/30						_		9/9	L
Thymus												
Hemorrhage			2/30					<u> </u>		<u> </u>	<u></u>	1/7
Stomach										1		
Dark red contents		ļ .	1/30	1/30					1			1/30

DOSE (mg/kg/d)		0		0		30		Schedu 20	84		46	80
DOSE (Ilig/kg/u/			t Purified diet		, ,	50		20	_ ا		١ '`	,00
Sex	М	F	M	F	М	F	M	F	М	F	M	F
Surviving												
Animals/group	25	24	Ð	0	28	24	29	23	25	20	0	0
Animals sacrificed	15	15	0	0	18	15	19	15	16	12	0	0
Adrenal gland						[
White foci		1/15	i		ļ <u>. </u>	1/15		2/15		1/12		
Adrenal gland						-	}					
Red foci	L	2/15	<u> </u>		1/18	3/15	1/19	4/15		2/12		
Heart		1	ŀ			1	1	ł	ŀ		ļ	1
White foci		<u> </u>	l			<u> </u>	<u> </u>		2/16			
Heart			ļ				1	1		1		l
Thickened		<u> </u>	<u> </u>			2/15						<u> </u>
Heart						1		1	İ	1		l
Red foci						1/15						<u> </u>
Kidney		1					ĺ	1	ŀ	1		1
White foci		.						1/15		1/12		L
Kidney												1
Cysts	1/15	<u> </u>			2/18	3/15	4/19	2/15	2/16	2/12		
Mammary gland												
Lactation		<u></u>	<u></u>			7/15		11/15		7/12		
Pituitary												
Enlarged		2/15				5/15	ł	4/15		1/12		

Testes		[·			<u> </u>	T	Γ	I		T		
Small	1	1	,			1	4/19		7/16		1	
Testes												
Soft		L]				3/19]	12/16			
Thymus]						
Hemorrhagic	<u> </u>	1/15				1.	2/19	2/15	l	1/12		
Uterus									_			
Red foci		L	l			1/15		1/15	İ	1/12		
Uterus		·						[
Cysts						1/15		3/15		1/12		
External surface												
Red matting		2/15			7/18	2/15	6/19	3/15	5/16	3/12		
Mass	j											
Subcutaneous		3/15	l		2/18	1/15		5/15	1/16			
Mass												
Submandibular	<u> </u>							1/15				
Tail, raised areas	Ì	i)			1				l i		
Multifocal	1/15	2/15			6/18	2/15	8/19	10/15	9/16	6/12		
Tail	1	1	i l				l	i		1	i	
Abscesses	2/15	1			1/18	1/15	1/19	1/15	1/16	2/12		
Lymph node	İ	ļ	1			l]	l			
Inguinal, enlarged	ļ	<u> </u>					4/19	1/15				
Lymph node	1	1						i				1
Inguinal, dark red						1/15		1/15		L		
Lymph node						1						- <u>- }</u>
Inguinal, green						ļ	5/19	ļ		2/12		
Fat												
Necrosis	1	L	Ĺl				1/19	1/15		L		

			of Anir	nals Sa				covery s				
DOSE (mg/kg/d)	1	0		9	11	B0	4	20	8-	40	16	80
		diet	Purifie	ed diet							L	
Sex	M	F	M	F	M	F	M	F.	M	F	M	F
Surviving												
Animals/group	10	9	0	0_	9	9	9	8	9	8	0	0
Animals sacrificed	10	9	0	0	9	9	9	8	9	8	0	0
Lymph node, Sub							1					
Reddened	l	2/9			1/9	3/9	1/9	1/8	5/9	2/8		
Adrenal glands]			1		
Red foci		1/9	1.		1/9	2/9	1/9	1/8	1/9	2/8		<u>L</u> .
Heart]	[·			I						
White foci							L		1/9	1/8		
Heart										Ī		
Enlarged		l							1/9	<u>L</u>	<u> </u>	
lleum												
Dark red contents									1/9			
Kidneys			İ	1 1		1	İ	ł		1		ł
Granular surface			<u> </u>		1/9	3/9	L	3/8	2/9			
Kidney, pelvis								! .	1	1	l	
Red streaks			<u> </u>		1/9	1/9		2/8				
Mammary glands									1		1	
Lactation		2/9				5/9		6/8		3/8	L	
Ovaries					-							
Cysts	`					3/9		1/8		_ 2/8		Ĺ
Ovaries										I		
White foci		2/9				5/9	L	4/8		5/8		L_
Testes												
Small	1/9				1/9		2/9		9/9]		
Mass												
Subcutaneous		1/9			1/9	3/9	1/9	3/8		<u> </u> ;		
Tail										-		

Raised areas			2/9	2/9	2/9	2/8	2/9	3/8	
Fat									
Necrosis							1/9		1 1

Histopathology: Incidence and severity.

DOSE (mg/kg/d)	0	ound Dead or S	180	420	840	1680
	diet	Purified diet				
Animals/group	30	30	30	30	30	30
Animals found dead/sacrificed	6	0	2	2	5	21
Cecum						1
Mucosal necrosis				<u> </u>		1/21(2)
Cecum, crypts				i		
Suppurative inflammation						1/21(1)
Cecum			•			2/21
Suppurative inflammation						1/21(2) 1/21(3)
Colon						
Dilatation of crypts						1/21(2)
Stomach		ļ - ·		i		4/21
Hyperkeratosis						3/21(2) 1/21(3)
Stomach Ulcer						2/21(2)
Epididymides				1	5/5	4/21
		{	1/2(3)			
Hypospermia			1/2(3)		1/5(3)	1/21(3)
Esophagus				 	4/5(4)	3/21(4)
				1/2/45		1
Hemorrhage Heart		 	·	1/2(4)		
Suppurative inflammation						1/21
Heart	•			1	3/5	1
Cardiomyopathy	1/6(2)		1/2(3)		1/5(2) 2/5(3)	
Heart				1/2/4)		
Atrial thrombus Kidney, Vacuolization,				1/2(4)		
cytoplasmic-tubular epithelium					1/5(3)	6/21(3)
Kidney, Mineralization				1 1		5/21
Tubular epithelium-medulla						2/5(1)
1						1/5(2)
						2/5(3)
Kidney, Protein casts					3/5	
Collecting tubules			1/2(2)	1/2(3)	2/5(2) 1/5(4)	1/21(1)
Kidney						2/24/2
Nephrosis		ļ			1/5(3)	2/21(3)
Kidney	•				415-51	
Cystic tubules				ļ	1/5(2)	1/21(2)
Kidney						
Nephropathy	1/6(1)		1/2(2)	1/2(4)		
Kidney					2/5	
Mineralization, pelvis					1/5(1) 1/5(2)	
Kidney					2/5	
Suppurative inflammation	·				1/5(3) 1/5(4)	
Liver						0/04/2:
Vacuolization, cytoplasmic						2/21(2)
Liver				1		

Liver Infiltrate, lymphocyte				1/5(2)	
Lungs . Edema, alveolar	3/6(2)	1/2(2)	1/2(2)	1/5(2)	3/21 1/21(2) 2/21(3)
Lungs Suppurative inflammation	1/6(2)				6/21 1/21(1) 3/21(2) 2/21(3)
Lungs Hemorrhage, alveolar	1/6(2)				2/21(2)
Lungs Perivascular edema	2/6(2)			2/5(2)	
Lympg node, Mesenteric Lymphoid atrophy				1/5(3)	1/21(4)
Rectum Suppurative inflammation		·			2/21 1/21(1) 1/21(2)
Spleen Lymphoid atrophy				1/5(3)	6/21 1/21(2) 5/21(3)
Testes Aspermatogenesis		1/2(3)		5/5 1/5(3) 4/5(4)	14/21 1/21(2) 10/21(3) 3/21(4)
Testes Multinucleated giant cells		1/2(2)			8/21 4/21(2) 4/21(3)
Testes Atrophy, seminiferous tubules		1/2(2)		4/5 3/5(3) 1/5(4)	2/21 1/2(2) 1/2(3)
Thymus Lymphoid atrophy				2/5(3)	14/21 4/21(2) 9/21(3) 1/21(4)
Thyroid gland Cytoplasmic vacuolization	·				6/21 1/21(2) 4/21(3) 1/21(4)
Tail Suppurative inflammation			1/2(2)	3/5 2/5(2) 1/5(3)	4/21 3/21(2) 1/21(3)

Histopathology of	Animals For	and Dead or S	acrificed I	Moribund (I	Females)	
DOSE (mg/kg/d)	0 diet	0 Purified diet	180	420	840	1680
Animals/group	30	30	30	30	30	30
Animals found dead/sacrificed	6	2	6	7	10	23
Cecum Mucosal necrosis					•	1/23(2)
Cecum, crypts Suppurative inflammation						9/23 4/23(1) 5/23(2)
Cecum Hemorrhage				·		2/23 1/23(1) 1/23(2)
Cecum Vasculitis						2/23 1/23(2) 1/23(3)

Cecum Submucosal edema				1/7(3)		3/23 1/23(2) 2/23(3)
Cecum Suppurative inflammation						5/23 3/23(2)
Color			ļ			1/23(3)
Colon Dilatation of crypts	 	. <u> </u>		<u> </u>		4/23 (2)
Colon Suppurative inflammation						9/23 1/23(1) 4/23(2) 3/23(3) 1/23(4)
Colon Necrosis						1/23(4)
Colon Submucosal edema						3/23 1/23(1) 2/23(2)
Stomach Hyperkeratosis						4/23(2)
Stomach			1	3/7		4/20(2)
Ulcer	1/6(3)			1/7(1) 2/7(2)		1/23(2)
lleum Villus atrophy						2/23(2)
Jejunum Villus atrophy						2/23(2)
Esophagus Suppurative inflammation	2/6(4)				1/10(4)	1/23(4)
Heart Suppurative inflammation	1/6(2)			1/7(4)		2/23 1/23(1) 1/23(2)
Heart Cardiomyopathy	1/6(2)			1/7(2)	1/10(1)	1/23(2)
Heart Necrosis, myocardial fiber						1/23(2)
Kidney, Vacuolization cytoplasmic-tubular epithelium						4/23(2)
Kidney, Mineralization Tubular epithelium-medulla		1/2(3)		1/7(3)	2/10 1/10(1) 1/10(2)	17/23 2/23(1) 10/23(2) 3/23(3)
Kidney, Protein casts						2/23(4)
Collecting tubules	1/6(2)		2/6(2)	1/7(2)	1/10(4)	
Kidney Nephrosis				1/7(2)		1/23(1)
Kidney Cystic tubules				.,,(2)	3/10 2/10(2) 1/10(3)	5/23 2/23(1) 3/23(2)
Kidney Nephropathy	1/6(3)			3/7 2/7(1) 1/7(4)	1,10(0)	0.20(2)
Kidney Mineralization, pelvis		·		2/7 1/7(2) 1/7(3)	6/10 5/10(2) 1/10(3)	
Kidney Suppurative inflammation				1/7(3)		

Liver					ſ	1
Vacuolization, cytoplasmic	1/6(3)		<u> </u>		1/10(1)	<u> </u>
Liver Necrosis, hemorrhage			-			2/23 1/23(2)
Liver Infiltrate, lymphocyte					1/10(1)	1/23(4)
Lungs Edema, alveolar	1/6(3)					
Lungs Suppurative inflammation	1/6(2)			2/7 1/7(2) 1/7(4)		2/23(2)
Lungs Hemorrhage, alveolar	1/6(2)	1/2(2)	2/6 1/6(2) 1/6(3)		1/10(2)	1/23(2)
Lungs Perivascular edema	3/6 2/6(2) 1/6(3)	1/2(2)			2/10 1/10(2) 1/10(3)	1/23(3)
Lymph node, Mesenteric Suppurative inflammation						2/23 1/23(2) 1/23(3)
Rectum Suppurative inflammation						5/23 1/23(1) 2/23(2) 1/23(3) 1/23(4)
Spleen Lymphoid atrophy				1/7(3)		3/23 2/23(2) 1/23(3)
Thymus Lymphoid atrophy	2/6 1/6(3) 1/6(4)			4/7 3/7(3) 1/7(4)	1/10(3)	11/23 3/23(2) 6/23(3) 2/23(4)
Thyroid gland Cytoplasmic vacuolization						4/23 2/23(2) 2/23(3)
Tail Suppurative inflammation				1/7(3)	2/10 1/10(3) 1/10(4)	4/23 2/23(2) 2/23(3)

Please note that only surviving animals in the purified diet control and HD groups were sacrificed at week 20.

His	topathology of Anima	als Sacrificed	at Week 20	
DOSE (mg/kg/d)	0 Purified diet	1680	0 Purified diet	1680
	MALI	S	FEMA	LES
Animals/group	30	30	30	30
Animals sacrificed	30	9	28	7
Colon Dilatation of crypts		1/9(4)		
Colon				
Suppurative inflammation	<u> </u>		<u> </u>	1/7(1)
Colon				4.77(2)
Lymphoid hyperplasia				1/7(3)
Parathyroid Follicular hyperplasia				1/7(2)
Epididymides				
Hypospermia	1/30(4)	8/9(4)		
Heart		•		
Cardiomyopathy	3/30(1)	1/9(1)	1/28(2)	

Heart Cell infiltrate, lymphocyte		1/9(2)		
Kidney, Mineralization	3/30	7/9	26/28	6/7
Tubular epithelium-medulla	1/30(1)	,,,		
rubular epitnellum-medulla		1/9(1)	2/28(1)	2/7(1)
	2/30(2)	5/9(2)	16/28(2)	2/7(2)
		1/9(3)	8/28(3)	2/7(3)
Kidney	1		5/28	
Cell infiltrate, lymphocyte	7/30(1)	1/9(2)	1/28(1)	
•	1		4/28(2)	
Kidney	3/30			
Nephropathy	2/30(1)		1/28(3)	
,	1/30(2)	•		
Kidney	1100		<u> </u>	
Hydronephrosis	1/30(2)		1	
Liver	1/30(2)		7/28	
	1		3/28(1)	477(2)
Necrosis	4/00/0\			1/7(2)
	4/30(2)		4/28(2)	
Liver	2/30		i i	
Vacuolization	1/30(1)	1/9(2)	1/28(1)	
	1/30(2)		·	<u> </u>
Liver	13/30	4/9		4/7
Cell infiltrate, lymphocyte	12/30(1)	3/9(1)	5/28(1)	2/7(1)
	1/30(2)	1/9(2)	1	2/7(2)
Pituitary				•
Adenoma, pars distalis	j		1/28P	
Pituitary			 	
Hyperplasia, pars distalis			1/28(2)	
Prostate	19/30	3/9	1,20(2)	
Suppurative inflammation	3/30(1)	1/9(1)	1	
Supporative inhamination			1 1	
	14/30(2)	2/9(2)	1	
	2/30(3)			
Rectum	1		i i	
Cell infiltrate, lymphocyte	<u> </u>	1/9(3)		1/7(2)
Rectum			1	
Нетоппаде			l l	1/7(1)
Siomach	14/30	6/9	5/28	
Suppurative inflammation	7/30(1)	1/9(1)	2/28(1)	
	7/30(2)	5/9(2)	3/28(2)	
Testes	1	8/9		
Aspermatogenesis	1/30(4)	4/9(3)]	
Aspenhalogenesis	1/30(7)	4/9(3) 4/9(4)	Į į	
Table 2	 			
Testes		9/9	 	
Edema, interstitial		5/9(2)]	
		4/9(3)		
Testes			Į	
Atrophy, seminiferous tubules	1/30(3)	7/9(3)		

Please note that only surviving animals in the ____ diet control, LD, MD and MHD groups were sacrificed at week 52. In addition, the epididymides, heart, kidneys, mammary glands, testes and thymus from all animals in the LD and MD groups were examined histologically at the week 52 sacrifice and the recovery (week 56) sacrifice.

·	listopathology of Anim	als Sacrificed	at Week 52	
DOSE (mg/kg/d)	0 diet	840	0 diet	840
	MALE	S	FEMA	LES
Animals/group	30	30	30	30
Animals sacrificed	15	16	15	12

Lymph node, submandibular		5/16	2/15	
Lymphoid hyperplasia	1/15(2)	2/16(2)	1/15(2)	
		3/16(3)	1/15(3)	
Parathyroid ·				
Follicular hyperplasia				1/12(2)
Adrenal gland				
Hyperplasia, medilla		1/16(2)		
Adrenal gland ·				
Pheochromocytoma		1/16P		
Eye			,	
Retinal regeneration		1/16(4)		
Eye				
Suppurative inflammation		1/16(3)		
Lungs				
Granulomatous inflammation				1/12(1)
Lungs				
Cell infiltrate, lymphocyte		1/16(1)		
Pancreas				
Vacuolization, cytoplasmic		1/16(2)		
Pituitary gland				
Adenoma	1/15P		4/15P	
Pituitary gland				
Hyperplasia, pars distalis		<u> </u>		1/12P
Stomach				• • • • • • • • • • • • • • • • • • • •
Dilatation, gastric glands	1/15(2)	4/16(2)		
Thyroid gland		2/16		. 1
Vacuolization, cytoplasmic		1/16(2)		
		1/16(3)		
Thyroid gland				
Follicular hyperplasia		2/16(3)		
Urinary bladder				
Suppurative inflammation				2/12(2)
Urinary bladder				-
Cell infiltrate, lymphocyte		1/16(3)		2/12(3)

For animals in the LD and MD groups, only the epididymides, heart, kidneys, mammary glands, testes and thymus from all animals were examined histologically at the week 52 sacrifice and the recovery (week 56) sacrifice.

	Histopa	thology o	f Animals	Sacrific	ed at wee	ek 52		
DOSE (mg/kg/d)	0 — diet		180		420		840	
Sex	M	F	M	F	M	F	М	F
Animals/group	30	30	30	30	30	30	30	30
Animals sacrificed	15	15	18	15	19	15	16	12
Epididymides Hypospermia			1/18(4)		3/19(4)		15/16 1/16(2) 14/16(4)	
Heart Cardiomyopathy	4/15 3/15(1) 1/15(2)	4/15 1/15(1) 3/15(2)	8/18 6/18(1) 2/18(2)	6/15 1/15(1) 515(2)	15/19 7/19(1) 5/19(2) 3/19(3)	12/15 8/15(1) 4/15(2)	11/16 1/16(1) 8/16(2) 2/16(3)	4/12 1/12(1) 3/12(2)
Kidney, Mineralization Tubular epithelium					1/19(4)	3/15 1/15(2) 2/15(3)	3/16 1/16(1) 2/16(2)	
Kidney Cell infiltrate, lymphocyte	3/15(1)	2/15(2)	12/18 1/18(1) 10/18(2) 1/18(3)	1/15(1)		4/15 2/15(2) 2/15(3)	5/16(2)	4/12 3/12(2) 1/12(3)

Kidney, Protein casts		5/15	8/18	12/15	5/19	5/15	7/16	7/12
Collecting tubules	1/15(1)	4/15(2)	1/18(1)	2/15(1)	2/19(1)	1/15(1)	1/16(1)	5/12(2)
Collecting tabules	1713(1)	1/15(3)	2/18(2)	8/15(2)	3/19(2)	2/15(2)	6/16(2)	2/12(3)
		17.10(3)	4/18(3)	2/15(3)	0,10(2)	2/15(3)	0/10(2)	2/12(3)
			1/18(4)	2 .0(0)		2.0(0)		
Kidney				3/15	4/19	7/15	2/16	7/12
Hyperplasia, pelvic	1/15(2))	2/18(2)	2/15(2)	1/19(1)	1/15(1)	1/16(1)	1/12(1)
epithelium	,	ļ		1/15(3)	3/19(2)	4/15(2)	1/16(2)	5/12(2)
i			ĺ	Į į		1/15(3)		1/12(3)
						1/15(4)		
Kidney	8/15	5/15	16/18	7/15	12/19	5/15	12/16	
Nephropathy	2/15(1)	1/15(1)	1/18(1)	5/15(2)	7/19(2)	1/15(1)	10/16(2)	
	6/15(2)	3/15(2)	10/18(2)	2/15(3)	2/19(3)	1/15(2)	1/16(3)	
		1/15(3)	5/18(3)		3/19(4)	2/15(3)	1/16(4)	
Vidani	<u> </u>		2/40	4/15	44/40	1/15(4)	0/46	7/40
Kidney Mineralization, pelvis		4/15(2)	2/18 1/18(2)	4/15 1/15(1)	11/19 3/19(2)	9/15 3/15(1)	8/16 1/16(1)	7/12 2/12(1)
willeralization, pelvis		4/15(2)	1/18(2)	2/15(2)	8/19(3)	5/15(1)	5/16(2)	5/12(1)
		 	1/10(3)	1/15(3)	0/15(3)	1/15(3)	2/16(3)	3/12(2)
Kidney		 	<u> </u>				2.5(5)	
Suppurative inflammation			2/18(2)	1/15(1)			2/16(2)	1/12(2)
Mammary gland				5/15		8/15		2/12P
Galactocele	,	2/15(2)	}	1/15(2)		2/15(2)		
				4/15(3)		6/15(3)		<u> </u>
Mammary gland						4/15		-
Active secretion			}	3/15(3)		3/15(2)		8/12P
		·				1/15(3)		
Testes				1	4/19	İ	14/16	
Aspermatogenesis			2/18(4)		1/19(3)		3/16(3)	
			42/40		3/19(4)	 	11/16(4)	
Testes	2/45/2)		13/18		11/19	İ	12/16	
Edema, interstitial	2/15(2)		3/18(1) 9/18(2)		4/19(1) 5/19(2)		2/16(2) 9/16(3)	
		,	1/18(4)		2/19(3)	l i	1/16(4)	
Testes			3/18		27.5(0)		15/16	
Atrophy, seminiferous			2/18(1)		4/19(3)		2/16(2)	
tubules			1/18(3)				12/16(3)	
						1	1/16(4)	
Testes								
Hyperplasia, interstitial			2/18(1)		1/19(1)]		
cell								
Thymus								
Lymphoid atrophy			<u></u>				1/16(3)	<u> </u>
Thymus		4145151	44544	3/15	446.5	,,,,,,,,		
Hemorrhage		1/15(2)	4/18(1)	1/15(1) 2/15(2)	1/19(1)	4/15(2)		
Thymus Thymoma							1/16	,
Tail			3/18		6/19	9/15	9/16	7/12
Suppurative inflammation	1/15(3)	1/15(3)	1/18(2)	2/15(3)	1/19(2)	3/15(2)	5/16(2)	1/12(2)
Cuppulative illianimation							J, . J . J,	
			2/18(3)		3/19(3)	5/15(3)	4/16(3)	5/12(3)

DOSE (mg/kg/d) .	die	t	18	0	42	20	8	40
Sex	M	F	M	F	М	F	М	F
Animals/group	30	30	30	30	30	30	30	30
Animals sacrificed	10	9	10	9	9	8	9	8
Epididymides		1						
Hypospermia .	1/10(4)		1/10(2)		3/9(4)		9/9(4)	
Heart	6/10	4/9	7/10		5/9	3/8	9/9	
Cardiomyopathy	2/10(1)	2/9(1)	2/10(1)	3/9(1)	1/9(1)	1/8(1)	7/9(1)	3/8(1)
• • •	4/10(2)	2/9(2)	5/10(2)	, ,	3/9(2)	2/8(2)	1/9(2)	` ` ′
	1	l			1/9(3)		1/9(3)]
Kidney, Mineralization		1	i				2/9	
Tubular epithelium, medulia		1/9(2)	1/10(2)	1/9(2)		1/8(2)	1/9(2)	2/8(3)
			<u> </u>				1/9(3)	` ′
Kidney, Hyperplasia	,					6/8	6/9	4/8
Pelvic epithelium	2/10(1)	1		2/9(2)	1/9(2)	- 2/8(1)	5/9(2)	3/8(2)
	ì		ļ			3/8(2)	1/9(3)	1/8(3)
			<u> </u>			1/8(3)		<u></u>
Kidney	7/10	5/9	10/10	5/9	8/9	6/8	4/9	2/8
Nephropathy	3/10(1)	2/9(1)	3/10(1)	1/9(2)	1/9(1)	1/8(1)	2/9(1)	1/8(2)
	4/10(2)	3/9(2)	4/10(2)	3/9(3)	6/9(2)	3/8(2)	1/9(2)	/1/8(3)
			3/10(3)	1/9(4)	1/9(3)	2/8(4)	1/9(3)	
Kidney, Mineralization	1	1	ł	i	3/9	5/8		6/8
Pelvis	1/10(2)	ŧ		3/9(2)	1/9(1)	2/8(2)	4/9(4)	4/8(2)
	ļ	}		1	1/9(2)	3/8(3)	i	2/8(3)
			 		1/9(3)		ļ	
Mammary gland		0,0,0	ł	5,0,0		4/8		0.0.0
Galactocele		2/9(3)		5/9(3)		1/8(2)		3/8(3)
			[2/8(3)		
			 			1/8(4)		
Mammary gland			ĺ			5/8		2/8
Active secretion	1	i e				4/8(2) 1/8(3)		1/8(2) 1/8(3)
Testes		 				1/0(3)		1/0(3)
Aspermatogenesis	[{		}	2/9(4)		9/9(4)	
Testes	2/10	 	7/10	 	5/9	 	9/9	ļ
Edema, interstitial	1/10(2)	1	3/10(1)	[2/9(1)		7/9(2)	
200mg, moronital	1/10(2)		3/10(1)		2/9(2)]	2/9(3)	
	1	1	1/10(3)		1/9(3)	1		
Testes	2/10		1				9/9	
Atrophy, seminiferous tubules	1/10(2)		1/10(2)		3/9(3)		2/9(2)	
	1/10(3)		1		(- /	1	7/9(3)	
Testes								
Hyperplasia, interstitial cell			2/10(2)		1/9(2)			
Tail			l				2/9	3/8
Suppurative inflammation	J		2/10(3)	1/9(3)	2/9(2)		1/9(2)	1/8(2)
••			` ` ′	`		!	1/9(3)	2/8(3)

Toxicokinetics:

Dose (mg/kg/d)		C _{max} (μg/ml)			T _{max} (hr)			AUC _{0-8hr} (μg.hr/ml)			
Males	Day 1	Week 11	Week 26	Week 52	Day 1	Week 11	Week 26	Week 52	Day 1	Week 11	Week 26	Week 52
180	3.81		3.03a	11.18	1.00	-	4.00a	1.00	16.39	•	b	40.21
420	5.98	·	16.50	16.44	0.50	1 -	0.50	1.00	30.89	-	62.85	76.75
840	6.39	-	20.23	28.70	0.50	1 -	1.00	1.00	39.39		86.30	84.65
1680	37.98	73.4		-	4.00	2.00		•	210.54	317.13	-	-

Females	Day 1	Week 11	Week 26	Week 52	Day 1	Week 11	Week 26	Week 52	Day 1	Week 11	Week 26	Week 52
180	4.60	-	2.47a	14.94	1.00		4.00a	0.50	16.02	•	Ь	32.76
420	7.33		23.80	14.83	1.00	-	0.50	1.00	33.55	•	64.75	56.14
840	13.50	-	32.50	45.03	0.50	•	1.00	1.00	58.40	-	96.77	99.62
1680	36.13	87.20	-	-	0.50	1.00			197.71		-	

^{- =} blood samples not collected.

Historical Incidence (%) and Severity of Myocardial Degeneration and Fibrosis Complex (Cardiomyopathy) in SD Rats (Modified from Ref. 3 i.e. Appendix 1).

		 -	Age ()	2 Morths)		
	Male	Female	Female			
		•		Grade		Grade
myocardial						
degeneration*	50	50	44	1.9	9	1.8
myocarditis*	50	50 .	60	2.0	16	1.8
nyocardial						
fibrosis*	50	50	31	2.2	11	1.8

The term myocardial degeneration and fibrosis complex is a combination of these histologic lesions.

Historical Incidence (%) and Severity of Cardiomyopathy in SD Rats in Searle Study SA3490.

		Age (13.5 months)								
Sex ·	Male	Female =		Male Grade		emale Grade				
SC-48334 dosages										
0 mg/kg	30	30	37	1.6	30	1.7				
180 mg/kg	30	30	53	1.6	30	1.6				
420 mg/kg	. 30	30	67	1.8	53	1.7				
840 mg/kg	30	30	77	1.9	27	1.4				

Summary of Study Findings:

In a 52-week chronic toxicity study in the rat with a 4 week recovery period, OGT 918 was administered by oral gavage (30/sex/group) at doses of 0, 180, 420, 840 and 1680 mg/kg/day (divided as three equal doses administered TID at 8 hr intervals). Due to high mortality in the 1680 mg/kg/day animals, dosing was terminated in this group during Week 10 and sacrificed during week 20. Clinical findings judged to be treatment-related at the lower dose levels consisted of tail findings in the 180, 420 and 840 mg/kg/day groups and soft stool in the 840 mg/kg/day group. Inhibition of bodyweight gain occurred in the 420, 840 and 1680 mg/kg/day groups throughout the treatment period. Inhibition of food consumption occurred in the 840 mg/kg/day males and 1680 mg/kg/day animals throughout the treatment period.

a = mean concentration values were not available at early time intervals (0.5 - 2 hr) and values reported for T_{max} and C_{max} may not be accurate.

b = sample quantity was not sufficient for reanalysis and concentration value was not available for AUC calculation. c = plasma concentrations were available only at 2 time-points and the AUC value was not used for data interpretation.

44/60 animals in the HD group (no AUC data at week 52) were found dead or sacrificed moribund during SC-48334 administration. Of the 44/60 animals found dead or sacrificed moribund, the cause of death or moribundity was not ascertained in 35/60 animals. Death of the remaining animals was attributed to septicemia, chronic renal disease, mechanical trauma, gavage error and enteropathy. 15/60 HMD (10x and 11x the maximum clinical dose of 100 mg TID based on AUC_{0-6hr} for males and females respectively) animals were found dead or sacrificed moribund during the study. The cause of death was not ascertained for 7/60 HMD animals. Death of the remaining animals was attributed to chronic renal disease, gavage error, bleeding technique and pituitary tumor. 9/60 animals in the MD group (9x and 6x the maximum clinical dose of 100 mg TID based on AUC_{0-6hr} for males and females respectively) were found dead or sacrificed moribund. The cause of death was not ascertained in 2/60 animals. Death of the remaining animals was attributed to gavage error, chronic renal disease, pituitary tumor and septicemia. 7/60 LD (5x and 4x the maximum clinical dose of 100 mg TID based on AUC_{BBb} for males and females respectively) animals were found dead or sacrificed moribund during the study. The cause of death was not ascertained in 6/60 animals. Death of the remaining animal was attributed to gavage error. 3/60 purified diet control females were found dead during the study. They were all females and the cause of death was not ascertained. In the diet control group, 11/60 animals were found dead or sacrificed moribund. The cause of death was not ascertained in 5/60 animals. Death of the remaining animals was attributed to gavage error and leukemia. Reviewer believes that demise of most of these animals was due to GI toxicity (stomach-ulcer, hyperkeratosis; cecum-mucosal necrosis, inflammation, hemorrhage colondilatation of crypts, necrosis, edema, inflammation; ileum and jejunum-villous atrophy), renal toxicity (nephrosis, nephropathy, vacuolation of tubular epithelium, inflammation, protein casts and mineralization of tubular epithelium and pelvis), hepatotoxicity (necrosis, cytoplasmic vacuolation, hemorrhage and lymphocytic infiltrate) and cardiac toxicity (cardiomyopathy, inflammation and necrosis of myocardial fiber) based on the histopathology data provided. All or most of the animals found dead had undergone autolysis. This might have confounded the histopathologic evaluation.

A dose-dependent decrease in % mean body weight was observed in both treated males and females. At 420 mg/kg/d (MD), the % decreases in mean body weight were 18% and 14% respectively for males and females. At 840 mg/kg/d (HMD), the values were 32% and 26% for males and females respectively. Percent decrease in food consumption also showed a dose-dependent effect. At 420 mg/kg/d, the % decreases in food consumption were 13% and 5% respectively for males and females. At 840 mg/kg/d, the values were 26% and 9% for males and females respectively. The decreased food consumption correlates with the decreased body weight.

Palpable ventral masses were observed in 7/60, 22/60, 23/60 and 18/60 animals in the control, LD, MD and MHD groups, respectively. No dose-relationship was apparent in either the total number of animals affected or the onset of occurrence at these dose levels. Only 2/60 animals in the HD group had palpable masses, however, this group was sacrificed early (during study week 20). Most were associated with mammary gland galactoceles or abscesses. Only 3 masses were found to be tumors; an adenoma was present in 1/60 LD animal and 2/60 control group animals each had a fibroadenoma. The other masses were abscesses or were associated with the mammary gland (lactation, galactoceles or active secretions).

Test article-related changes in hematology parameters, serum chemistry parameters and urinalysis were observed at all dose levels. After the recovery, all hematology values, serum chemistry values and urinalysis values were comparable to those in the control group.

Equatorial cataracts were observed in treated males relative to controls. The incidence appears to be dose-related. In treated males the incidence are 1/28 (LD), 1/29 (MD) and 18/27 (HMD) at week 52. In treated females, the incidence at the HMD dose is 9/23 at week 52. After the 4 week recovery period, the incidence had decreased to are 1/10 (LD), 1/9 (MD) and 5/9 (840 mg/kg) in treated males and 4/8 for treated females. In the HD group that was terminated at week 20, the incidence of cataracts was 9/9 (males) and 5/7 (females) at week 14. The cataracts may have been induced by the pharmacologic activity of the drug. OGT 918 causes perturbations in lipid metabolism. Perturbations in lipid metabolism are associated with lipid peroxidation, which generates free radicals. Oxygen free radicals are important known causes of tissue damage including cataracts.

Gross and microscopic findings indicated that the testis, epididymides, GI tract, eyes, heart and kidney were the principal test-article affected organs. Soft and/or small testes were seen grossly at LD, MD, HMD and HD. Apparently treatment and dose-related increases in aspermatogenesis, interstitial edema, and atrophy of seminferous tubules were observed microscopically in the LD, MD, HMD and HD animals. The incidence and severity of the lesions were essentially unchanged 4 weeks after cessation of treatment. Cardiomyopathy and nephropathy were the principal pathologic changes seen in the heart and kidneys in control, LD, MD and HMD animals found dead or sacrificed during the course or at termination of the study. The incidence of the heart and kidney findings increased with dose. Sponsor stated that these changes were considered to be species- or age-related. Literature supporting sponsor's claim is summarized in appendix 2.

The target organs of toxicity include the epididymides (hypospermia), kidney (nephropathy, lymphocyte infiltration, protein casts, hyperplasia and mineralization), testes (atrophy of seminiferous tubules, aspermatogenesis, edema, and hyperplasia of interstitial cells), GI tract (stomach-ulcer, hyperkeratosis; cecum-mucosal necrosis, inflammation, hemorrhage colon-dilatation of crypts, necrosis, edema, inflammation; ileum and jejunum-villous atrophy), eyes (cataracts), mammary gland (galactocele, active secretion) and tail (suppurative inflammation). Most of these toxicities occurred at all dose levels and showed very little/no recovery at the end of the treatment free period. NOAEL could not be established since toxicities were observed at the LD tested.

Study title: 52-Week Gavage Toxicity Study with SC-49483 in Cynomolgus Monkeys with 8 weeks of recovery.

Key study findings: KEY STUDY FINDINGS:

- 10 animals (5/24 control; 1/12 LD; 4/24 HD) died or were sacrificed in a moribund condition during this study; however, none of the deaths or unscheduled sacrifices were considered to be related to the test material. 8 of the 10 unscheduled deaths were directly related to sequelae of the gastric catheterization, most commonly inflammation and infection at the skin or stomach implantation site and peritonitis. Of the remaining two animals in the control group, one had significant inflammation of the large intestine, while the cause of moribundity of the other was not determined.
- The test material produced dose-dependent increases in fecal changes (white feces, yellowish substance in feces and organ changes during treatment. At the end of the recovery period, the fecal changes were still present in addition to red feces.
- QRS interval was slightly but statistically significantly increased by 0.02 sec in HD males relative to control. At the end of the recovery period, this parameter was comparable to that of control.

- RBC, HGB and HCT were slightly but statistically significantly decreased in HD females relative to control. Reticulocytes were statistically significantly increased by 50% in HD females relative to control. Prothrombin time was slightly but statistically significantly increased in HD females relative to control. At the end of the recovery period, RBC was still slightly but statistically significantly decreased in HD females. Platelets were statistically significantly increased in HD males by 136% relative to control. Basophils were slightly but statistically significantly decreased in HD males relative to control.
- Creatinine was slightly but statistically significantly decreased in the HD group relative to control. Total protein and chloride levels were also slightly but statistically significantly decreased in HD males. At the end of the recovery period, albumin was slightly but statistically significantly decreased in HD males whereas phosphate and bile acids were statistically significantly increased in HD females by 138% and 267% respectively relative to control. The increased bile acids may be suggestive of impaired liver function and cholestasis.
- Urine phosphorus was statistically significantly decreased by 71% in HD males relative to control. At the end of the recovery period, urine chloride excretion and urine calcium were statistically significantly decreased by 33% and 58% respectively in HD males and females.
- Mean sperm concentrations were highest for males in the control group (7.73 x 10⁸), and decreased for males given 750 and 2000 mg/kg/day (4.02 x 10⁸ and 2.96 x 10⁸, respectively). This corresponds to 48% and 62% decrease in the LD and HD males. The number of amorphous sperms was 1% in HD males compared to zero for control.
- T4 was statistically significantly decreased in LD females by 32% and in HD males and females by 41% and 37% respectively relative to control. At the end of the recovery period, T4 was still statistically significantly decreased in HD females.
- The target organs of toxicity include the GI tract cecum, colon, stomach (pigmented macrophages, granulomatous inflammation), Liver (pigmented macrophages, vacuolation), Pancreas (↓ zymogen granule staining), adrenal gland (mineralization), thyroid gland (vacuolated macrophage), seminal vesicle (mineralization), lymph nodes mesenteric & submandibular (granuloma, mineralized, pigmented macrophage), mammary gland (lymphocytic infiltrate), skin (acanthosis/hyperkeratosis), cervix (lymphocytic infiltrate), skeletal muscle (inflammation, necrosis/degeneration), brain (mineralization, necrosis), spinal cord (mineralization) and kidney (angiectasis-glomerulus).
- NOAEL could not be established because of lesions in the liver, pancreas, seminal vesicle, brain and skeletal muscle at the LD.
- At the end of the recovery period, the following lesions were observed mostly in the HD group Pancreas (↓ zymogen granule staining), submandibular salivary gland (atrophy, mineralization), Stomach (inflammation, fibrosis), heart, seminal vesicles, thyroid & parathyroid (lymphohistiocytic infiltrate), adipose tissue (fibrosis, inflammation, pigmented macrophage), cervix (mucopurulent exudate), spleen (fibrosis), skin (acanthosis/hyperkeratosis, fibrosis, crusting, ulceration), brain (mineralization), spinal cord (demyelination, axonal swelling) and kidney (tubular pigment, membranoproliferative glumerulopathy).

Study no: SA 4078

Volume #, and page #: Vol. 35, pg. 1. Conducting laboratory and location:

Date of study initiation: August 12, 1993. GLP compliance: Yes (U. S. A. and U. K.)

QA- Report: Yes (X) No ()

Drug, lot #, radiolabel, and % purity: Drug lot # 93K003-B2A; radiolabeled SC-49483 LOT #s GDS-2448-146, GDS-2448-148, GDS-3768-45, GDS-3768-47, GDS-3768-64, GDS-3768-65. 100% pure.

Formulation/vehicle: A solution of SC-49483 in a carrier made up of 0.5% methylcellulose (w/v), and 0.1% polysorbate 80 (v/v) in reverse osmosis water.

Methods (unique aspects):

Dosing: SC-49483 was administered at total daily doses of 750 and 2,000 mg/kg/d for 52 weeks. The total daily dose was divided into 3 doses/day each separated by 8 hours. The drug was administered through a catheter surgically implanted into the lumen of the stomach through week 19. After week 19, the catheters were tied off and animals were dosed by oral gavage up to week 52. A surgical control group (did not undergo any surgical or dosing procedures) and a vehicle control group were included in the study.

Species/strain: Monkey/Cynomolgus.

#/sex/group or time point (main study): Control: 12 animals/sex/group; 750 mg/kg/day: 6 animals/sex/group; 2000 mg/kg/day: 12 animals/sex/group.

Satellite groups used for toxicokinetics or recovery: 12 animals/sex/group (0 mg/kg/d); 6 animals/sex/group (750 mg/kg/day); 12 animals/sex/group (2000 mg/kg/day) for TK. 6 animals/sex/control and HD group were designated as recovery animals for 8 weeks after the 52 week treatment period.

Age: Young adult Weight: 2.5 to 7.0 kg.

Doses in administered units: 750, 2000 mg/kg/day.

Route, form, volume, and infusion rate: gavage via intragastric catheter; 10 ml/kg/dose.

•		Dose		
CTOUT	(mg/kg/day) ⁶	Concentration [mg/mL]	<u>Humber</u>	of Animals Person
1 (Control)	D	D	12 ^b	125
2 (Lov) c	750	25.0	6	6
3 (Righ) ^e	2000	66.7	126	12 ^b
4 (Instrument	M.V.	MY.	4	4

- a. The dose volume was 10 ml/kg/dose. When administering [¹⁴C] SC-49483, the volume administered was adjusted to maintain the nominal dose of μci/kg. Each animal received the daily dosage divided into three doses/day. The control group received the carrier only.
- b. Six animals/sex were designated as recovery animals and continued without treatment for at least 8 weeks after at least 52 weeks of treatment.
- c. Three animals/sex/group in groups 2 and 3 received [14C] SC-49483 once on Day 1 and once each during Weeks 26 and 52.
- d. The animals in Group 4 were maintained in the jacket and tether system but did not undergo the surgical procedure nor were dosed with test material or carrier.

Observations and times:

Clinical signs: Twice daily.

Body weights: Recorded prior to treatment, on day 1 of treatment and weekly thereafter.

Food consumption: Daily.

Ophthalmoscopy: Was conducted before initiation of treatment; during Weeks 4, 8, 12, 26, and 52; and during Weeks 56 and 60 (recovery animals).

EKG: Conducted before initiation of treatment; during Weeks 13, 26, 39, and 52; and during Week 56 (recovery animals).

Hematology: Blood was collected from all animals. Animals were fasted for ~ 24 hours prior to blood collection. Blood was collected twice before initiation of treatment (pre- and post-surgery), during weeks 16, 32 and 52; and during weeks 56 and 60 (recovery animals).

Clinical chemistry: The blood collected for hematology evaluation was also used for routine clinical chemistry evaluation including hormone analysis.

Urinalysis: Urine samples were collected for approximately 24 hours for routine urinalysis evaluation.

Gross pathology: Organs/Tissues collected for gross pathology examination is indicated in the list of addendum.

Organs weighed: Organs weighed are indicated in the addendum list.

Histopathology: Tissues collected for histopathology examination is indicated in the list of addendum.

Toxicokinetics: Blood samples were collected on Day 1 and during Weeks 26 and 52 at approximately 0.5, 1, 3, 5, and 8 hours after the first daily dose. For [14C] SC-49483, blood samples were collected at approximately 0.5, 1, 3, 5, 8, and 24 hours after the [14C] SC-49483 dose. Urine and feces were collected for 168 hours after each radioactive dose at approximately 24-hour intervals. Urine was collected into containers surrounded by wet ice.

Other:

Sperm evaluation: At scheduled sacrifice, sperm was collected from the left epididymis from each male animal and evaluated for concentration, motility and morphology.

Hormone analysis: Serum was harvested from blood samples collected during weeks 16, 32 and 52; and during weeks 56 and 60 from recovery animals. TSH, T3, T4 and testosterone levels were measured.

Results:

Mortality:

		MO	RTALITY			
	Vehicle	Control	750 mg	/kg/day	2000 mg	g/kg/day
Sex	M	F	М	F	M	F
Incidence	2/12	3/12	1/6	0/6	2/12	2/12
Total	5/	24	1/	12	4/	24

Ten animais (5 males and 5 females) died or were sacrificed in a moribund condition during this study. However, none of the deaths or unscheduled sacrifices were considered to be related to the test material. 8 of the 10 unscheduled deaths were directly related to sequelae of the gastric catheterization, most commonly inflammation and infection at the skin or stomach implantation site and peritonitis. Of the remaining two animals in the control group, one had significant inflammation of the large intestine, while the cause of moribundity of the other was not determined.

Clinical signs: Incidence of clinical signs during weeks 1-53. Rectal body temperature and

respiration rate were unremarkable during the treatment and recovery periods.

Dose (mg/kg/d)	0 (vehicle)		7	750		2000		0 (No surgery nor vehicle)	
Sex	M	F	M	F	M	F	M	F	
White feces		2/12	4/6	4/6	12/12	12/12			
Yellowish substance in feces		1/12	3/6	1/6	12/12	12/12			
Red skin (abdomen)	1/12			1/6		3/12	1/4		
Red skin (hind limbs)				1/12	3/12	2/12			
Skin (broken-L hind limb)		1/12	2/6	2/6	4/12	2/12			
Red nasal discharge	6/12	4/12	3/6	1/6	8/12	6/12	I		
		RE	COVERY (WEEKS 53-	61)				
Red feces				I .		2/12			
White feces					4/12	4/12			
Yellowish substance in feces					4/12	4/12			

Empty cells indicate zero incidence

Body weights: No treatment-related effects on body weight. At the end of the treatment period, mean body weight of treated animals and those of controls were not statistically significantly different.

Food consumption: No treatment-related effects.

Ophthalmoscopy: Unremarkable.

Electrocardiography:

Dose (mg/kg/d) Week 52	0 (veh	icle)	75	Ò	2000		0 (No surgery not vehicle)		
Sex	M	F	M	F	M	F	M	F	
Heart rate (beats/min)	167		175		160		T		
•	18.9		29.5		27.6			l	
P-R interval (sec) M	0.08		0.08		0.09				
SD	0.000		0.000		0.019		l	<u></u>	
QRS interval (sec)	0.03		0.03		0.05*			T	
	0.008		0.009		0.013		•		
Q-T interval (sec)	0.21		0.21		0.22				
	0.030		0.027		0.034]		
			RECOVERY (WEEKS 5	6)				
At the end of the recover	y period the	prolonge	ed QRS interva	al (HD mal	es) had returne	d to norm	al limits.		

* p< 0.05

Hematology:

WEEK 52 DATA

			• • •	,,						
Dose (mg/kg/d)	0 (ve	hicle)			50			2000		surgery nor vehicle)
Sex	M	F	М		F		М	F	М	F
RBC (E6/UL)	6.0	09 ± 0.369		5.0	5 ± 0.	513		5.29 ± 0.264*		6.25 ± 0.580
HGB (g/dl)	1	1.1 ± 0.71	· ·	10).6 ± 0	.50		10.2 ± 0.81*		11.4 ± 0.73
HCT (%)	30	6.3 ± 2.17		34	.8 ± 1	.75		33.5 ± 2.31°		37.5 ± 2.47
RETIC (%)	0	.2 ± 0.18		0	.2 ± 0.	13		0.4 ± 0.15*		0.2 ± 0.10
PT (sec)	1	0.9 0.29	T	10	0.6 ± 0	.29		11.4 ± 0.43°		10.8 ± 0.26
		,	RECOV	/ERY	(WEE	KS 60)			
Dose (mg/kg/d)	0	(vehicle)		7	50		-	2000	0 (ñ	o surgery nor vehicle)
Sex	M	F		M	F		M	F	M	F
PLT (E3/UL)	380 ± 77.	5				518	± 62.4*			
BASO (E3/UL)	0.1 ± 0.4					0.0	± 0.0*			
RBC (E6/UL)		6.26 ± (0.434					5.40 ± 0.606*		

^{*} p< 0.05

Clinical chemistry:

WEEK 52 DATA

Dose (mg/kg/d)	0 (vei	hicle)	750 2000		00	0 (No surgery no vehicle)		
Sex	M	F	M	F	M	F	М	F
Creat (mg/di)	1.1 ± 0.16	7.9±0.30	1.2 ± 0.15	7.8±0.49	1.0 ± 0.11*	7.3±0.51*		
T. Pro (g/dl)	8.0 ± 0.58	±	7.7 ± 0.43	±	7.4 ± 0.68*	±		<u> </u>
Chol (mg/dl)	136±14.4	±	103±17.4*	±	105±18.8*	±		
			RECOVERY	(WEEKS 60))			
Dose (mg/kg/d)	0 (vel	nicle)	75	60	20	00		rgery nor icle)
Sex	M	F	M	F	M	F	M	F
ALB (g/dl)	4.5±0.26				4.0±0.26*			
I PHOS (mg/dl)		5.5±0.75				7.6±0.84*		
Bile acids (umol/l)		3.0±2.0				8.0±2.6*		

Urinalysis:

URINE CHEMISTRY WEEK 52 DATA

Dose (mg/kg/d)	0 _. (ve	hicie)	750	1	200	2000		surgery ehicle)
Sex	M	F	M	F	M	F	M	F
U PHOS (mg/dl)	21 ± 14		11 ± 11.5		6 ± 6.0°			
	•		RECOVERY (NEEKS (50)			
Dose (mg/kg/d)	0 (ve	hicle)	750)	200	00		surgery ehicle)
Sex	M	F	M	F	M	F	M	F
CL EXCR (mmol)	7.8 ± 1.29		±		5.2 ± 0.87°			
U Ca (mg/dl)	,	15.9±4.22		+		6.6±4.83°		

* p< 0.05

Organ weights: No statistically significant differences in absolute or relative organ weights at the terminal or recovery sacrifices.

Gross pathology:

WEEK 52 DATA (TERMINAL SACRIFICE)

Dose (mg/kg/d)	0 (ve	hicle)	75	50	200	00	0 (No	surgery ehicle)
Sex	M	F	M	F	M	F	M	F
Lung				1				2.5
Red foci		7/4		1/6	1/6	1/6	<u> </u>	L
Adhesions	2/5	1/4	1/5	1/6	4/6			,
Stomach			7				1	
Adhesions	0/5	2/4	3/5	5/6	2/6	2/6	1	<u> </u>
Dark foci	1/5]				
Thickened wall			1		<u> </u>	1/6		
Cecum						T		
Dark foci			1/5		1/6	<u> </u>	L	
Colon			T				1	
Dark foci						1/6	<u> </u>	
Ovary					<u> </u>			
Cyst(s)				1/6		1/6	J	
Small				1/6				
		RECC	VERY SACE	IFICE (WEE	KS 60)			
Dose (mg/kg/d)	0 (ve	hicle)	7.	50	20	00		surgery ehicle)
Sex	M	F	M	F	М	F	M	F
Liver			1					
Red foci					7/4		<u> </u>	
Spleen								
Opaque capsule		1/5				2/4		1

GROSS PATHOLOGY (UNSCHEDULED DEATHS)

Dose (mg/kg/d)	0 (ve	hicle)	75	50	2000	
Sex	M	F	M	F	M	F
N	2	3	1	0	2	2
Lung Red foci			1/1			
Diffusely light	l	1/3				
Dark foci		1/3				
Kidney Thickened capsule		1/3			1/2	
Dark foci	1/2		l			
Heart Red foci					1/2	
Liver						

Light foci		1/3		7		
Mottled		1/3	I			
Spleen						
Large		2/3	1			
Mottled		1/3				
Granular surface		1/3	1			
Stomach						
Red foci						1/2
Catheter site-stomach		1				
Thickened	1/2	<u>L</u>	1/1	j		2/2
Diffusely red	1/2		1/1			1
Red foci		1				1/2
Submandibular L. node						
Large		İ	ŀ	1	1/2	
Peritoneum/cavity				1		
Adhesions		1/3	ļ	l l	1/2	<u> </u>
Pleura/cavity		l .		•		
Adhesions		L	Ī			
Thickened		1/3			1/2	

Sperm evaluation:

Dose (mg/kg/d)	0 (vehicle)	750	2000
Sperm concentration (108)	7.73 ± 11.14	4.02 ± 5.41	2.96 ± 3.97
Sperm morphology			
Amorphous	0.0 ± 0.0	0.0 ± 0.4	1.0 ± 1.6

a = number of sperms/gram of caudal tissue

Hormone analysis:

WEEK 52 DATA

Dose (mg/kg/d	1)	0 (vehicle)		750		2000		T	0	
, , ,								(No surger	y nor vehicle)	
Sex		M	F	M	F	M	F	M	F	
Testosterone (ng/dl)		212	32.0	242	23.1	223	32.8		32.8	
•	•	285	9.52	157	2.66	182	12.9	1	13.44	
TSH (µU/ml)	M	3.2	3.4	2.7	3.2	3.0	2.90		2.9	
.,	SD	0.57	0.81	0.23	0.80	0.92	0.30	1	0.55	
T4 (μg/dl)	М	3.4	4.1	3.6	2.8*	2.0*	2.6*		4.2	
	SD	0.58	0.61	0.92	0.56	0.46	0.61	1	0.34	
T3 (ng/dl)		No data	was prov	ided for T3.						

WEEK 56 DATA

Dose (mg/kg/d)	Testosterone (ng/dl)		TSH (µU/ml)		T4		T3	
Sex	M	F	M	F	M	F	M	F
0 mg/kg/d	77.5	34.2	4.4	4.5	3.8	3.9	111	95.8
	13.33	8.35	0.91	0.31	0.77	0.64	24.2	11.64
2000 mg/kg/d	203	41.5	4.3	3.8*	2.9	3.1	87.8	116
	227.2	6.36	1.24	0.45	0.42	0.65	25.84	16.7

* p< 0.05

WEEK 60 DATA (RECOVERY)

Dose (mg/kg/d)	Testosterone (ng/dl)		TSH (μU/ml)		T4		Т3	
Sex	M	F	M	F	M	F	М	F
0 mg/kg/d	126	30.7	4.1	4.1	3.9	3.50	140	122
	73.3	9.57	0.77	0.38	1.05	0.42	24.9	23.6
2000 mg/kg/d	131	45.9	4.1	3.6	2.9	2.6*	111	110
	106.2	22.03	1.23	0.50	0.80	0.68	28.0	32.7

* p< 0.05

Histopathology:

WEEK 52 I	DATA	TERMINAL	SACRIFICE)

Dose (mg/kg/d)	0 (ve	ehicle)	7	50	2000		
Sex	M	F	M	F	M	F	
Cecum		T	1		2/6		
Pigmented macrophage	1		}	·	1/6(1)		
	j	1		ł	1/6(2)		
Colon]		
Pigmented macrophage -	- 1	1			1/6(2)		
Liver						2/6	
Pigmented macrophage	1/5(1)	ł	1		1/6(1)	1/6(1)	
, ,	` ` `	1	j '		` '	1/6(2)	
Vacuolation		1	1/5(2)	1/6(2)	1/6(1)		
Pancreas			5/5	5/6	6/6	6/6	
↓ Zymogen granule staining			4/5(1)	4/6(1)	4/6(1)	4/6(1)	
-,	İ	j	1/5(2)	1/6(2)	2/6(2)	2/6(2)	
Stomach	· ·						
Granulomatous inflammation	j	1	1	1/6(1)		1/6(1)	
Adrenal gland							
Mineralization		}	ì		1/6(2))	
Thyroid						j	
Vacuolated macrophage	i '	1				1/6(1)	
Cervix							
Lymphocytic infiltrate	1	1	j i		ı	1/6(1)	
Seminal vesicle		 	2/5			,	
Mineralization	1	}	1/5(1)		1/6(1)		
		1	1/5(2)				
Mesenteric L. node	······						
Granuloma, mineralized	1	i	1		1/6(1)		
Pigmented macrophage		· · · · · · · · · · · · · · · · · · ·	1			1/6(1)	
Submandubular L. node			ļ · · · · · · ·				
Pigmented macrophage	1	}	1			1/6(1)	
Mammary gland			1			3/6	
Lymphocytic infiltrate	İ	ł.	1			2/6(1)	
_,,,	[ļ				1/6(2)	
Skin		1	1				
Acanthosis/hyperkeratosis	ļ					1/6(1)	
Skeletal muscle					2/6		
Subacute inflammation	ŀ	ļ		1/6(1)	1/6(1)	,	
, , , , , , , , , , , , , , , , , , ,	· 1	•		(.,	1/6(2)	1/6(2)	
Necrosis/degeneration		1/4(1)	1/5(1)	2/6	2/6		
	· }] " ('')	1	1/6(1)	1/6(1)	1/6(1)	
		1		1/6(2)	1/6(2)	""	
Brain			 				
Mineralization, vascular]		1/5(1)	Ì	1/6(1)		
Mineralization			1/5(1)		2/6(1)	1/6(1)	
Necrosis, white matter			1/5(1)				
Spinal cord			1				
Mineralization, vascular	1		j 1		1/6(1)		
Kidney							
Angiectasis, Glomerulus)		1/6(1)		

RECOVERY SACRIFICE (WEEK 61)							
Pancreas							
↓ Zymogen granule staining		3/4(1)					
Submandibular salivary gland							
Atrophy	1/4(1)						
Mineralization	1/4(1)	1/4(1)					

Stomach						
Chronic active inflammation			1	1	1/4(2)	
Fibrosis, serosal			I		1/4(2)	
Heart						
Lymphohistiocytic infiltrate			1	<u>l</u> l .	1/4(1)	
Parathyroid			T .			
Lymphohistiocytic infiltrate	1		L		1/4(1)	
Thyroid						
Lymphohistiocytic infiltrate				<u> </u>		1/4(1)
Adipose tissue						
Fobrosis	J]	1 1		1/1(3)
Pigmented macrophage						1/1(2)
Chronic active inflammation						1/1(2)
Cervix			1			
Mucopurulent exudate	<u> </u>			1		1/4(3)
Seminal vesicles						
Lymphohistiocytic infiltrate			-	1	1/4(1)	
Spleen	T :		T			2/4
Capsular fibrosis	1/5(1)	1/5(1)	i	1 1		1/4(2)
·	<u> </u>		<u> </u>			1/4(3)
Skin	Τ –					
Fibrosis, subcutaneous	<u> </u>		<u> </u>		1/4(2)	
Acanthosis/hyperkeratosis					1/4(2)	•
Superficial crusting					1/4(3)	
Ulceration			<u> </u>		1/4(3)	
Brain			<u> </u>			
Mineralization		1/5(1)		<u> </u>	2/4(1)	
Spinal cord						
Demyelination/Axonal swelling			1			1/4(1)
Kidney						
Tubular pigment			<u> </u>			1/4(1)
Membranoproliferative glumerulopathy			<u> </u>			1/4(3)

1 = minimal, 2 = slight, 3 = moderate, 4 = marked, 5 = severe

UNSCHEDULED SACRIFICE

Dose (mg/kg/d)	0 (vehicle)		750		2000	
Sex	M	F	M	F	М	F
Cecum						
Inflammation, lymphohistiocytic	<u> </u>	1/3(2)	<u> </u>			
Inflammation, chronic active	1/2(3)					
Colon						
Inflammation, lymphohistiocytic		1/3(2)	L			
Inflammation, chronic active	1/2(3)					
Jejunum						
Dilatation, lymphatic vessel	1/2(2)					
Liver	1		1 1		2/2	
Lymphohistiocytic infiltrate	1/2(1)	2/3(1)	1/1(1)		1/2(1)	1/2(1)
					1/2(1)	
Vacuolation, centrilobular		<u> </u>	<u> </u>		1/2(1)	
Necrosis	1/2(2)		1			
Suppurative inflammation		1/3(4)				
Chronic active inflammation		1/3(4)				
Hepatocellular degeneration		1/3(2)				
Fibrosis		1/3(2)				
Pancreas						_
Lymphohistiocytic infiltrate		2/3(1)	<u> </u>		<u> </u>	
Vacuolation		1/3(1)				
↓ zymogen granules			1/1(1)			
Parotid salivary gland	1/2(1)	2/3				
Lymphohistiocytic infiltrate		1/3(1)	<u> </u>		1	

F		1/3(2)	1		1
Submandibular salivary gland		1/3(2)			
Lymphohistiocytic infiltrate	l l	1/3(2)			ŀ
Stomach		1/3(2)	 		
		4/2/2\			1
Chronic active inflammation		1/3(3)	 		
Erosion, junction of esophagus &	i	4/0/0			Ī
stomach		1/3(2)	Į		ļ
Heart	ļ				
Hemorrhage			ļ	1/2(1)	
Lymphohistiocytic infiltrate		1/3(1)		<u> </u>	
Adrenal gland			[1
Extramedullary hematopoiesis		1/3(2)			<u></u>
Thyroid gland			1 1		1
Lymphohistiocytic infiltrate	1/2(1)				
Skin - catheter site	Ì		1	·	2/2
Inflammation, chronic active	1/2(3)	1/3(3)	1/1(4)	1/2(2)	1/2(2)
			li		1/2(4)
Inflammation, granulomatous			1/1(1)	1/2(2)	
Fibrosis	1/2(2)	2/3			
	`	1/3(2)]]	1/2(2)	1
1	1 1	1/3(3)) 1	_, ,	1/2(3)
Нетопраде	1		1/1(2)		1/2(2)
Bacterial colony		1/3P	1		2/2P
Acanthosis/hyperkeratosis		1/3(3)			
Stomach - catheter site	1	.,0,0,	 	<u> </u>	
Lymphohistiocytic infiltrate			j 1	1/2(1)	ال.
Chronic active inflammation –			├──┤	1/2(1)	2/2
submucosa/muscularis	1/2(3)	2/3(3)	1/1(3)	1/2(3)	1/2(3)
Suumuusamustulans	1/2(3)	ചാര്യ	"'(3)	1/2(3)	
Hamashana (aubar 1995)	4/0/0	 -	4/4/2		1/2(4)
Hemorrhage (submucosa)	1/2(2)	2/2/2)	1/1(3)	4/0/4)	
Foreign material	1/2(2)	2/3(3)	1/1(2)	1/2(1)	000
Bacterial colony			ļ		2/2P
Hemorrhage (mucosa)		L	ļI		1/2(1)
Fibrosis		1/3(3)	Ļ———		ļ
Pericardial sac	1 1		j	•	!
Lymphohistiocytic infiltrate		1/3(2)			
Edema		1/3(3)	<u> </u>		L
Peritoneal cavity			J - 7		1
Inflammation, granulomatous	1/2(3)	1/3(4)	<u> </u>	1/2(4)	L
Pleural cavity					
Inflammation, granulomatous			<u> </u>	1/2(5)	!
Fibrosis				1/2(1)	
Vagina					
Lymphohistiocytic infiltrate		3/3(2)	[2/2(1)
Axillary lymph node					<u> </u>
Lymphoreticular hyperplasia) [1/3(4)) i		}
Lumbar/iliac lymph node	1		 		
Lymphoreticular hyperplasia		1/3(4)	1		1
Lymphocytic hyperplasia					1/2(2)
Hemorrhage					1/2(1)
Mesenteric lymph node	 		 		
Edema	<u> </u>	1/3(2)	(]	
		1/3(2)	 		
Lymphoid depletion		113(2)	 		1/2/1\
Hemorrhage	4/2/2		} 		1/2(1)
Germinal centers, decreased	1/2(2)		 		
Submandibular lymph node	1 1				4,0,0
Hemorrhage			 		1/2(2)
Depletion, lymphoid		1/3(2)			
Renal lymph node	•			1	
Hemorrhage		<u> </u>	<u></u> _	·	1/2(2)
Mesentery			L I		